

In-vivo Antioxidant Effects of the Orally Administered Paracetamol, Aqueous Extracts of *Salvia triloba*, and *Origanum syriacum*

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Abstract

The plant extracts are found to be effective against oxidative stress. There are lots of reasons for increased oxidative stress in which environmental factors and life style play a big role. In this study the herbal extracts of *Salvia triloba*, and *Origanum syriacum* used to study antioxidant properties compared to paracetamol. A total of 27 volunteers were divided into 3 groups, 2 groups took 250 ml of aqueous extract of *S. triloba*, and *O. syriacum* for 5 days and 3rd group took paracetamol 500 mg, each tablet twice a day for five days. Blood specimens were collected before taking extract and one hour after taking the first dose at day one and one day after the last dose. Serum Total Antioxidant Status (TAS) and reduced glutathione (GSH), malondialdehyde (MDA) and Superoxide Dismutase (SOD) in RBCs were tested. The oral intake of aqueous extract of the plants for five days increased serum TAS and GSH in RBCs compared to zero time of intake. In addition, oral intake of aqueous extracts of *Salvia triloba* for five days increased significantly, P value ≤ 0.05 the activity of SOD in RBCs as compared to zero time of administration. The herbs extracts of *Salvia triloba* and *Origanum syriacum* have anti-oxidant property compared to paracetamol, so as medicinal herbs with antioxidant effects can inhibit produced oxidative effects.

Keywords: Antioxidant status, Malondialdehyde, Reduced glutathione, Superoxide dismutase

Introduction

There are lots of reasons for increased oxidative stress in which environmental factors and life style play a big role. Antioxidants can stop the harmful consequences of oxidative stress on human health (Valko *et al.*, 2006; 2007).

Medicinal plants are potential sources of natural antioxidants (AS *et al.*, 2020). Natural antioxidants can inhibit the oxidative damages through metal chelating properties and scavenging activity of free radicals (Soobrattee *et al.*, 2005); in addition, plant extracts are effective against oxidative stress.

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Salvia triloba and *Origanum syriacum* are the most popular medicinal plants in Lebanon and Jordan that has been commonly used for medical purposes (Barakat & Fatima, 2000).

Salvia triloba, belongs to the Lamiaceae family, commonly referred to as Greek sage, the Arabic Name of *Salvia triloba* is Meramiyyh. It is commonly used to treat gastric disorders, bloating, infections of the mouth, gums, and teeth, stomach discomfort, headaches, common cold, cough, and influenza (Barakat & Fatima, 2000). *Salvia triloba* extracts have anti-inflammatory, antioxidant, antimicrobial and anticancer activities (Kamatou *et al.*, 2005; 2007; 2008; 2010; Añides *et al.*, 2019).

Origanum syriacum L. (Lamiaceae), is often known as white oregano and/or Syrian oregano, and its Arabic name is za'atar (Seidemann, 2005). Previous studies have reported its acetylcholinesterase inhibition and antioxidant activity (Loizzo *et al.*, 2009, Zein *et al.*, 2011); antifungal agent detection (Abou-Jawdah, 2004; Kintzios, 2004); antibacterial activity (Kintzios, 2004; Peña *et al.*, 2019) analgesic activity, expectorant, antirheumatic, sedative, anthelmintic and antiparasitic activities (Zein *et al.*, 2011).

All over the world, paracetamol is an analgesic and antipyretic drug, and single-component and multi-component preparations can be obtained without a doctor's prescription.

In vitro antioxidant properties of *Salvia triloba* and *Origanum syriacum* extracts were previously evaluated (Bilto *et al.*, 2015c). This study proposed to see the antioxidants *in vivo* effects of the orally intake of aqueous extracts of *S. triloba* (leaves), *O. syriacum* (leaves), and Paracetamol.

Materials and Methods

Plant Material

Plant materials: Meramiyyh (leaves), oregano (leaves) were bought from an herbal shop in Madaba city. The herbs are selected according to its wide use as a traditional medicine.

Formation of Aqueous Extracts

250 gram of *S. triloba*, *O. syriacum* dried and was boiled in 11.250 Liter water for 15 min, and left dousing for 4 hours at 25 °C. Then



the aqueous extract was packed in tidy bottles with a capacity of 1.250 L.

Blood Samples

27 participants were divided into three groups of nine persons, and their demographic data is presented in **Table 1**. The two groups drank 250 ml of water extracts of selected medicinal plants (Meramiyyh, Oregano) 3 for 5 days and the third group took paracetamol (500 mg, each tablet twice a day) for 5 days.

Three blood samples were collected (one before taking water extract, one after 1st dosage taking water extract on day one, and one day after the last dose) in the gel clot activator tube. To separate and collect the serum, the gel tube was centrifuged at 3000 rpm for 10 minutes at 25°C. Then, 2 ml of deionized water was added to the cells in the tube under the gel, the tube was centrifuged at 3000 rpm for 5 min, and the supernatant was collected. For further testing, all specimens were stored at -20 °C.

Table 1. Demographic Data of the Participants

	Age	Sex
	Mean ± S.D.	(Female/Male)
<i>O. syriacum</i>	35.8±14.7	4/5
<i>S. triloba</i>	42.8±14.6	6/3
Paracetamol	30.6±9.8	3/6

Serum TAS Assay

TAS analysis of serum was performed using Randox TAS kit. The data is expressed in millimoles/Liter.

MDA Activity in RBC

MDA was measured as a marker of lipid peroxidation using Dormandy's technique (1971) and Thiobarbituric Acid (TBA) with minor modifications. All MDA values were shown in nmol/gHb units.

GSH Activity in RBC

Ellman's technique was slightly modified to analyze red blood cell GSH (Ellman, 1951). All GSH values are expressed in mg/gHb.

SOD Activity in RBC

SOD levels were measured using a Randox kit. The outcomes were displayed as U/gHb.

Statistical Analysis

Mean and standard deviation were analyzed using SPSS software, version.17 to evaluate test data.

Results and Discussion

In vivo study showed that oral intake of *S. triloba* and *O. syriacum* increased serum TAS after 5th day of extract intake. Paracetamol did not affect serum total antioxidant capacity (**Table 2**).

Oral administration of *S. triloba* and *O. syriacum* also resulted in an increase in GSH in RBCs at 5th day of extract intake. Paracetamol had no effect on GSH in RBCs (**Table 3**). However, SOD in RBCs increased considerably after five days of oral intake of *S. triloba* extract in comparison to 0 time of intake, whereas, *O. syriacum* and Paracetamol did not increase erythrocyte activity of superoxide dismutase (**Table 4**).

Salvia triloba, *Origanum syriacum* and Paracetamol did not decrease erythrocyte content of MDA after five days of administration (**Table 5**).

Table 2. TAS Before and After Oral Intake of the Studied Herbs

	TAS (mmol/l)		
	0 time	1hr (day 1)	Day 6
<i>Salvia triloba</i>	1.12±0.11	1.16±0.15	1.22±0.16*
<i>Origanum syriacum</i>	1.14±0.10	1.21±0.11	1.28±0.09*
Paracetamol	1.4±0.27	1.36±0.3	1.31±0.9

* Each statistic reflects the mean± S.D., (n=9), with a *P value of ≤ 0.05 when in comparison to the 0 time administration.

Table 3. GSH Measured in mg/g Hb Before and After Oral Intake of Studied Herbs

	GSH	
	0 time	Day 6
<i>Salvia triloba</i>	0.54±0.09	0.87±0.10*
<i>Origanum syriacum</i>	0.73±0.11	0.80±0.10*
Paracetamol	0.73±0.12	0.75±0.13

* Each statistic reflects the mean ± standard deviation (n =9), with a *P value of ≤ 0.05 in comparison to the 0 time administration.

Table 4. SOD Measured in U/gHb Before and After Oral Intake of Studied Herbs

	0 time	Day 6
	<i>Salvia triloba</i>	868.0±167.1
<i>Origanum syriacum</i>	1037.1±155.3	1098.0±181.5
Paracetamol	1114.5±256.6	1091.2±172.1

* Each statistic reflects the mean± standard deviation (n =8), with a *P value ≤ 0.05 in comparison to the 0 time administration.

Table 5. MDA Measured in nmol/g Hb Before and After Oral Intake of Studied Herbs

	MDA	
	0 time	Day 6
<i>Salvia triloba</i>	17.9±3.4	15.8±3.8
<i>Origanum syriacum</i>	17.5±3.4	16.4±3.5
Paracetamol	16.1±3.1	17.8±4.1

* Each number reflects the mean ± standard deviation (n =9).

Oral intake of aqueous extracts of *S. triloba* and *O. syriacum* enhanced the serum TAS after five days of intake. This conclusion is supported with the results of other studies (Bilto & Alabdallat, 2015a; 2015b; Alabdallat, 2016; 2021). Our present study also presented that the plants increased GSH content of RBC after five days of intake. This conclusion is consistent with the results of other studies (Bilto & Alabdallat, 2015a; 2015b; Alabdallat, 2016; 2021) and (El-Dakhakhny *et al.*, 2000; Ali, 2004; Kaleem, *et al.*, 2006; EL-shenawy *et al.*, 2008; Asnani & Verma, 2009) who showed that oral treatment of ginger or nigella extract increased GSH or TAS in kidney and liver tissues of rats and mice. However, the results also indicated that SOD content of RBC increased after five days of *Saliva triloba* intake as also reported in studies of (Bilto & Alabdallat, 2015a; 2015b; Alabdallat, 2021) and (Ajith *et al.*, 2007; Bakirel *et al.*, 2008; Asnani & Verma, 2009). They demonstrated that oral rosemary or ginger extract can significantly enhance Superoxide Dismutase (SOD) activity in mouse liver or diabetic rabbit serum. The current investigation also found that the tested plants did not reduce the red blood cell content of MDA after five days of treatment. This conclusion is supported the results of similar studies.

Conclusion

As our findings are based on volunteers with no history of oxidative stress, we can suggest the use of herbal extracts of plants like *Saliva triloba*, *Origanum syriacum* as medicinal herbs with antioxidant effects can improve the basal defense mechanism for oxidative stress, and this may help in decreasing oxidative stress related disease.

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Conflict of interest: None

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Ethics statement: This research was approved by the Jordan University Committee and therefore meets the ethical standards mentioned in the 1964 Declaration of Helsinki. Before participating in the study, each individual provided informed permission.

References

Abou-Jawdah, Y., Wardan, R., Sobh, H., & Salameh, A. (2004). Antifungal activities of extracts from selected Lebanese wild plants against plant pathogenic fungi. *Phytopathologia Mediterranea*, 43(3), 377-386.

Ajith, T. A., Hema, U., & Aswathy, M. S. (2007). Zingiber officinale Roscoe prevents acetaminophen-induced acute hepatotoxicity by enhancing hepatic antioxidant status. *Food and Chemical Toxicology*, 45(11), 2267-2272.

Alabdallat, N. G. (2016). In vivo antioxidant related effects of orally administered aqueous extract of lemon balm (*melissa officinalis* L.) in human. *International Journal of Pharma and Bio Sciences*, 7(3), 642-645

Alabdallat, N. G. (2021). Antioxidant Properties of Orally Administered of Aqueous Extracts of Selected Medicinal Plants and Paracetamol in Human Volunteers: In vivo. *Journal of Contemporary Medical Sciences*, 7(3), 179-182.

Ali, B. H. (2004). The effect of Nigella sativa oil on gentamicin nephrotoxicity in rats. *The American Journal of Chinese Medicine*, 32(01), 49-55.

Añides, J. A., Dapar, M. L. G., Aranas, A. T., Mindo, R. A. R., Manting, M. M. E., Torres, M. A. J., & Demayo, C. G. (2019). Phytochemical, antioxidant and antimicrobial properties of the white variety of 'Sibujing' (*Allium ampeloprasum*). *Pharmacophore*, 10(1), 1-12.

AS, A., Nayeem, N., MA, A., & Imran, M. (2020). Antimicrobial and antioxidant screening of the solvent extracts of the leaves and stem of sesuvium portulacastrum. *Pharmacophore*, 11(4), 5-10.

Asnani, V. M., & Verma, R. J. (2009). Ameliorative effects of ginger extract on paraben-induced lipid peroxidation in the liver of mice. *Acta Poloniae Pharmaceutica*, 66(3), 225-8.

Bakirel, T., Bakirel, U., Keleş, O. Ü., Ülgen, S. G., & Yardibi, H. (2008). In vivo assessment of antidiabetic and antioxidant activities of rosemary (*Rosmarinus officinalis*) in alloxan-diabetic rabbits. *Journal of Ethnopharmacology*, 116(1), 64-73.

Barakat, A. R., & Fatima, A. (2000). Treatment with medicinal plants in Jordan. *Dirasat. Medical and Biological Sciences*, 27(1), 53-74.

Bilto, Y. Y., & Alabdallat, N. G. (2015). Ex vivo and In vivo antioxidant related effects of zingiber officinale roscoe (Ginger) extracts in humans. *European Journal of Medicinal Plants*, 99-108.

Bilto, Y. Y., & Alabdallat, N. G. (2015). In vitro and in vivo antioxidant related effects of rosemary (*Rosmarinus officinalis* L.) extracts in humans. *American Journal of Clinical and Experimental Medicine*, 3(5), 213-221.

Bilto, Y. Y., Alabdallat, N. G., & Salim, M. (2015). Antioxidant properties of twelve selected medicinal plants commonly used in Jordan. *Journal of Pharmaceutical Research International*, 121-130.

El Shenawy, N. S., Soliman, M. F., & Reyad, S. I. (2008). The effect of antioxidant properties of aqueous garlic extract and Nigella sativa as anti-schistosomiasis agents in mice. *Revista do Instituto de Medicina Tropical de São Paulo*, 50(1), 29-36.

El-Dakhakhny, M., Barakat, M., Abd El-Halim, M., & Aly, S. M. (2000). Effects of Nigella sativa oil on gastric secretion and ethanol induced ulcer in rats. *Journal of Ethnopharmacology*, 72(1-2), 299-304.

Ellman, G. L. (1951). Tissue Sulfhydryl (-SH) Groups. *Archive of Biochemistry and Biophysics*, 82(1), 70-77.

Kaleem, M., Kirmani, D., Asif, M., Ahmed, Q., & Bano, B. (2006). Biochemical effects of Nigella sativa L seeds in diabetic rats. *Indian Journal of Experimental Biology*, 44(9), 745-748

- Kamatou, G. P. P., Viljoen, A. M., Gono-Bwalya, A. B., Van Zyl, R. L., Van Vuuren, S. F., Lourens, A. C. U., Başer, K. H. C., Demirci, B., Lindsey, K. L., Van Staden, J., et al. (2005). The in vitro pharmacological activities and a chemical investigation of three South African Salvia species. *Journal of Ethnopharmacology*, 102(3), 382-390.
- Kamatou, G. P., Van Vuuren, S. F., Van Heerden, F. R., Seaman, T., & Viljoen, A. M. (2007). Antibacterial and antimycobacterial activities of South African Salvia species and isolated compounds from *S. chamelaeagnea*. *South African Journal of Botany*, 73(4), 552-557.
- Kamatou, G. P., Van Zyl, R. L., Davids, H., Van Heerden, F. R., Lourens, A. C. U., & Viljoen, A. M. (2008). Antimalarial and anticancer activities of selected South African Salvia species and isolated compounds from *S. radula*. *South African Journal of Botany*, 74(2), 238-243.
- Kamatou, G. P., Viljoen, A. M., & Steenkamp, P. (2010). Antioxidant, antiinflammatory activities and HPLC analysis of South African Salvia species. *Food Chemistry*, 119(2), 684-688.
- Kintzios, S. E. (2004). *Oregano: The Genera Origanum and Lippia*. 1st ed. Taylor & Francis.
- Loizzo, M. R., Menichini, F., Conforti, F., Tundis, R., Bonesi, M., Saab, A. M., Statti, G. A., de Cindio, B., Houghton, P. J., Menichini, F., et al. (2009). Chemical analysis, antioxidant, antiinflammatory and anticholinesterase activities of *Origanum ehrenbergii* Boiss and *Origanum syriacum* L. essential oils. *Food Chemistry*, 117(1), 174-180.
- Peña, J. F., Dapar, M. L. G., Aranas, A. T., Mindo, R. A. R., Cabrido, C. K., Torres, M. A. J., Manting, M. M. E., & Demayo, C.G. (2019). Assessment of antimicrobial, antioxidant and cytotoxic properties of the ethanolic extract from *Dracontomelon dao* (Blanco) Merr. & Rolfe. *Pharmacophore*, 10(2), 18-29.
- Seidemann, J. (2005). *World Spice Plants: Economic Usage, Botany, Taxonomy*. 1st ed. Berlin, Heidelberg: Springer-Verlag.
- Soobrattee, M. A., Neergheen, V. S., Luximon-Ramma, A., Aruoma, O. I., & Bahorun, T. (2005). Phenolics as potential antioxidant therapeutic agents: mechanism and actions. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 579(1-2), 200-213.
- Valko, M., Leibfritz, D., Moncol, J., Cronin, M. T., Mazur, M., & Telser, J. (2007). Free radicals and antioxidants in normal physiological functions and human disease. *The international Journal of Biochemistry & Cell Biology*, 39(1), 44-84.
- Valko, M., Rhodes, C. J. B., Moncol, J., Izakovic, M. M., & Mazur, M. (2006). Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chemico-Biological Interactions*, 160(1), 1-40.
- Zein, S., Awada, S., Rachidi, S., Hajj, A., Krivoruschko, E., & Kanaan, H. (2011). Chemical analysis of essential oil from Lebanese wild and cultivated *Origanum syriacum* L.(Lamiaceae) before and after flowering. *Journal of Medicinal Plants Research*, 5(3), 379-387.