Investigating the Effects of Diabetes Mellitus on Several Biochemical Parameters and Histopathological Changes of Some Organs in Rats

C. Delgado-Montemayor*, P. Cordero-Pérez, R. Salazar-Aranda, N. Waksman-Minsky

Received: 24 February 2024 / Received in revised form: 01 May 2024, Accepted: 09 May 2024, Published online: 15 May 2024

Abstract

One of the metabolic diseases that occurs frequently in elderly women is diabetes. In this study, the effects of diabetes on some biochemical parameters and histopathological changes in rats were investigated. In this experimental-interventional study, adult female rats were divided into four groups of 8. The first control group, the second ovariectomy group (for 55 weeks), the third induced diabetes group in the last 5 weeks of the study in mice with ovaries, and the fourth ovariectomy group (for 55 weeks) and induced diabetes in the last 5 weeks of the study. Pathological changes in lung, pancreas, kidney, and liver tissues and some serum biochemical indices were determined in each group at the end of the study. Diabetic and ovariectomized diabetic rats revealed an increase in blood glucose, alkaline phosphatase, alanine aminotransferase, gamma-glutamyl transferase, aspartate aminotransferase, urea, high-density lipoprotein, calcium, and triglyceride compared to the control group (p < 0.05). Ovariectomized rats showed an increase in cholesterol, lowlipoprotein, high-density lipoprotein, dehydrogenase, and triglyceride compared to the control group (p < 0.05). The findings revealed that the long-term reduction of estrogen in ovariectomized diabetic rats can have beneficial changes in reducing the serum levels of glucose, aspartate aminotransferase, alanine aminotransferase, urea, gammaglutamyl transferase, very low-density lipoprotein, and triglyceride compared to non-ovariectomized diabetic rats.

Keywords: Diabetes mellitus, Biochemical parameters, Histopathological changes, Rats

Introduction

Diabetes Mellitus is one of the metabolic diseases in humans and animals that due to the decrease in insulin secretion from the beta

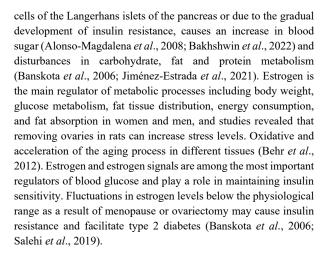
$\begin{array}{lll} C. & Delgado-Montemayor^*, & R. & Salazar-Aranda, & N. \\ Waksman-Minsky & & & \\ \end{array}$

Department of Analytical Chemistry, Medical School, Autonomous University of Nuevo Leon, Monterrey, Mexico.

P. Cordero-Pérez

Liver Unit at the "Dr. José E. González" University Hospital of the Autonomous University of Nuevo Leon, Monterrey, Mexico.

*E-mail: cdelgado.me0018@uanl.edu.mx



Recently, it has been reported that estrogen receptors alpha and beta regulate glucose transport by glucose transporter 4 (GLUT4) and cause tissue sensitivity to insulin (Alonso-Magdalena *et al.*, 2008; Sekido *et al.*, 2019). Also, the short-term reduction of estrogen in ovariectomized rats causes changes in circulating insulin levels and sensitivity of insulin receptors, and this disorder in glucose homeostasis after eight weeks of ovariectomy in rats with significant changes in lipid profiles such as There was an increase in serum cholesterol, triglyceride, low-density lipoproteins and non-esterified fatty acids and a significant decrease in high-density lipoproteins, but it is not clear what effect postmenopausal estrogen has on carbohydrate metabolism (Zhang *et al.*, 2002).

The role of ovarian hormones in glucose metabolism and insulin sensitivity is not well known (D'Eon *et al.*, 2002; Liebmann *et al.*, 2022). However, in a study, a decrease in insulin sensitivity was reported in the middle of the luteal phase compared to the middle of the follicular phase of the menstrual cycle and during the normal pregnancy period (Horton *et al.*, 2002). Short-term use of estrogen replacement hormones in postmenopausal women with diabetes improved the level of glycosylated hemoglobin (Andersson *et al.*, 1997). However, studies by Ferrara *et al.* did not show a difference between the levels of glycosylated hemoglobin in diabetic women treated with estrogen compared to untreated women (Ferrara *et al.*, 2001; Tanaka *et al.*, 2017).

Also, Godsland *et al.* (1993) studies showed that estrogen does not change fasting glucose levels, insulin levels, and increased insulin



sensitivity in women after menopause. In a study, it was found that long-term use of estrogen in postmenopausal women may increase the risk of type 2 diabetes (Lau *et al.*, 2021).

Okada et al. studies revealed that there is no significant difference in the level of glycosylated hemoglobin in Paleo women treated with replacement hormones compared to women who did not use replacement hormone therapy (Okada et al., 2003; Speksnijder et al., 2023). Due to the change in sedentary lifestyle, nutrition, genetics, etc., the incidence of diabetes has increased in society, and for women, diabetes may also occur with increasing age in addition to menopause. Despite the mentioned fact, little research has been done on the effect of diabetes on blood sugar, lipid profiles, and liver and kidney enzyme indices in long-term menopause. Therefore, the current study aimed to determine the impact of experimentally induced diabetes on blood sugar, lipid profiles, and enzyme indices of liver, kidney, and histopathological changes of pancreas, liver, and kidney in ovariectomized rats with long-term reduction of estrogen.

Materials and Methods

This experimental-interventional study was done on 32 female Wistar rats with an age range of 12 to 14 weeks and an average weight of 200 ± 200 grams over 55 weeks.

Study Groups, Ovariectomy, and Diabetes Induction in Rats

Female rats were randomly divided into 4 groups of eight (Dogru et al., 2012). Then, before surgery, female rats were anesthetized with a combination of xylazine and ketamine drugs at a dose of 80 and 7 mg per kilogram of body weight, respectively, intramuscularly, and after shaving the hair on the back surface of the back and cleaning with ethanol, shearing 20 mm length was created on the surface of the skin and 5 mm diagonal incisions were made on the muscles of the left and right testicles. Surgery to remove the ovaries was performed (Sadeghi et al., 2009). To induce experimental diabetes in ovariectomized and nonovariectomized rats, a subcutaneous injection of alloxan (Sigma Aldrich, made in Germany) in the amount of 150 mg per kilogram of body weight in a physiological serum solution was performed. To make better conclusions and to deal with the competitive effect of glucose with alloxan, which might prevent the definitive effect of the drug on the beta cells of the pancreas, the intended injection was performed in each of the rats at 8:00 am and in the fasting state. After 72 hours of alloxan injection, a blood sample was provided from the tail vein of each rat, and their blood glucose levels were measured by a Glucocard made in Japan and their blood sugar was above 200 mg/dL. The criterion was their becoming diabetic (Unal et al., 2011).

In the first group, the surgical control group, the fat surrounding the ovary and the ovary was directed out and returned to its place, and the skin of the back surface of the waist was sutured with non-absorbable thread (Sadeghi *et al.*, 2009) and for 55 weeks under standard conditions. In the second group, the ovariectomy group was kept for 55 weeks after surgery and removal of the ovary. The third group was the group of rats with ovaries that were kept for 55 weeks and became diabetic with alloxan in the last 5 weeks of the research period. The fourth group is the ovariectomy group and in

the last 5 weeks of the research period, diabetics were treated with alloxan injection and kept under standard conditions.

Determination of Serum Enzymes

At the end of the 55th week, the mice were anesthetized with chloroform after fasting for 12 hours while they had access to water, and after opening the chest cavity, blood was taken from the heart. Blood was collected in tubes without anticoagulant and serum was separated by an H-11N centrifuge manufactured by KOKUSAN, Japan at 3000 rpm for 10 minutes. Serum blood sugar, AST (aspartate aminotransferase), ALT (alanine aminotransferase), creatinine, urea, **CPK** (creatinephosphokinase), LDH (lactate-dehydrogenase) enzymes were measured by auto analyzer model 3000-BT manufactured by Biotecnica, Italy. Was performed. Serum concentrations of triglycerides, low-density lipoproteins, total cholesterol, and highdensity lipoproteins were measured using recommended guidelines. Also, the amount of very low-density lipoproteins (VLDL) was determined using the Friedewald formula (Warnick et al., 1990) and plasma atherogenic index, cardiac risk, and atherogenic coefficient (Ikewuchi et al., 2014).

To study the tissue changes, immediately after blood collection, the tissues were placed in 10% buffered formalin for fixation, and according to the usual histological methods, the blocks of Paraffin were prepared and 5-micron thick sections were cut with a microtome and stained with hematoxylin and eosin in the common method.

Statistical Method

Based on the data collected using SPSS version 23 statistical software, the mean \pm standard deviation was calculated. Data analysis was done with one-way analysis of variance and if there was a significant difference, the differences between the groups under study were compared using Tukey's test at a significant level of P < 0.05.

Results and Discussion

The amount of blood glucose in female diabetic rats was significantly higher than in diabetic ovariectomized rats (P = 0.008). The serum activity of gamma-glutamyltransferase, alanine aminotransferase, and aspartate aminotransferase was increased in female diabetic rats compared to rats in the surgical control group (P < 0.05). Also, serum alanine aminotransferase activity was increased in female diabetic rats compared to ovariectomized rats (P = 0.036). The amount of serum albumin in ovariectomized diabetic and diabetic rats was lower than that in rats of the surgical and ovariectomy control groups (P = 0.05). Alkaline phosphatase enzyme in female diabetic rats and ovariectomized diabetic rats showed a significant increase compared to the surgical control group (P < 0.05). Lactate dehydrogenase enzyme in ovariectomized and ovariectomized diabetic rats showed a significant increase compared to the surgical control group (P = 0.001) and the diabetic rat group (P = 0.04). Creatine phosphokinase enzyme in ovariectomized diabetic rats was more than the surgical control group (P = 0.001). The percentage ratio of liver weight to body weight in ovariectomized rats showed a significant decrease compared to the surgical control group (P = 0.026). Also, the percentage ratio of liver weight to body weight in the ovariectomized diabetic and diabetic rats group was more than in the ovariectomized rats group (P = 0.05).

The amount of serum calcium in the group of diabetic and ovariectomized rats had a significant decrease compared to ovariectomized rats (P = 0.000). However, no significant change was observed compared to the surgical control group (P = 0.828). Also, the amount of serum calcium in ovariectomized diabetic rats

showed a significant decrease compared to the sham group (P = 0.000). The amount of serum phosphorus in the ovariectomy group, diabetic ovariectomy group, and diabetic group did not show a statistically significant difference compared to the surgery control group (P > 0.01). Body weight in ovariectomized rats was higher than in surgery control rats, diabetic rats, and diabetic ovariectomized rats (P = 0.002). The serum concentration of urea in female diabetic rats was higher than surgical control group and ovariectomized non-diabetic rats (P = 0.003) (Table 1).

Table 1. Comparison of changes in serum urea, creatinine, and average kidney weight in the groups under study

Groups Biochemical parameters	Control	Ovariectomized female rat	Diabetic rat	Ovariectomized and diabetic female rats	P-value
Urea (mg/dl)	67.500±3.138°	66.83±2.959°	224±58.20a**	145.600 ± 16.154^{b}	0.001
Creatinine (mg/dl)	0.859±0.128 ^a	1.083 ± 0.116^a	1.100±0.108 ^a	$0.960{\pm}0.081^a$	0.248
Percentage of weight of kidneys/body weight	0.749±0.018 ^a	0.614±0.033ª	1.065±0.077 ^a	0.922±0.109 ^a	0.300

Values are Mean ± SD. The number of samples in each group is 6 mice, non-similar letters (a, b, c) in each view of the table indicate a significant difference at the 95% confidence level.

The concentration of serum creatinine in the rats of the experimental groups did not show a statistically significant difference compared to the surgical control group (P = 0.01) (Table 1). The percentage ratio of kidney weight to body weight in diabetic female rats was more than the surgical control group (P = 0.022). Also, the average ratio of kidney weight to body weight in diabetic and ovariectomized rats showed a significant increase compared to the ovariectomized group (P = 0.001) (Table 1). The amount of serum triglyceride in ovariectomized rats and diabetic

rats showed a significant increase compared to the group of control rats (P < 0.01). Also, the number of triglycerides in diabetic female rats was more than that of the ovariectomized group (P = 0.002). However, the amount of triglyceride in ovariectomized diabetic rats was significantly lower than in diabetic rats (P = 0.000). The amount of serum cholesterol in ovariectomized rats was significantly higher than the surgical control group (P = 0.032) (Table 2).

Table 2. Comparison of fat profiles in the studied groups.

Biochemical parameters	Groups Control	Ovariectomized female rat	Diabetic rat	Ovariectomized and diabetic female rats	P-value
Triglyceride (mg/dl)	42.833±3.33°	104.500±9.704 ^{b**}	182.250±24.294 ^{a***}	$84.00{\pm}7.463^{b}$	0.000
Cholesterol (mg/dl)	53.66±5.684 ^b	84.33±7.596 ^a	63.750±4.697 ^{ab}	69.600±8.58 ^{ab}	0.053
HDL cholesterol (mg/dl)	42.43±7.681 ^b	87.300±12.107 ^a	46.775±6.144 ^b	51.42±6.48 ^b	0.008
LDL cholesterol (mg/dl)	19.833±2.243 ^b	37.316±4.260 ^{a*}	25.275±1.409 ^b	36.794±5.718 ^{a*}	0.027
VLDL (mg/dl)	8.56±0.666°	20.90±1.940 ^b	36.45±4.857 ^a	16.8±1.492 ^b	0.000

Values are Mean ± SD. The number of samples in each group is 6 mice, non-similar letters (a, b, c) in each view of the table indicate a significant difference at the 95% confidence level.

The amount of high-density lipoprotein in the group of ovariectomized rats revealed a significant increase compared to the surgical control group (P = 0.016) (**Table 2**). The amount of low-density lipoprotein in ovariectomized rats and diabetic ovariectomized rats revealed a significant increase compared to the surgical control group (P = 0.01) (**Table 2**). Very low-density lipoproteins in diabetic rats revealed a significant increase

compared to ovariectomized rats (P=0.002) and rats of the surgical control group (P=0.000). Also very low-density lipoproteins in ovariectomized rats, it was significantly higher than the surgical control group (P=0.007) (Table 2). The ratio of triglycerides to high-density lipoproteins in diabetic and ovariectomized diabetic rats was higher than in the surgical control group (P=0.01) (Table 3).

^{**} Statistically significant difference compared to the control group (P<0.01)

^{**} Statistically significant difference compared to the control group (P<0.01)

^{***} Statistically significant difference compared to the control group (P<0.001)

Table 3. Comparison of atherogenic indices in different groups under study.

•	_		· .			
Biochemical parameters	Groups	Control	Ovariectomized female rat	Diabetic rat	Ovariectomized and diabetic female rats	P-value
TG/HDL		1.186 ± 0.204^{b}	1.259 ± 0.136^{b}	$3.947 \pm 0.359^{a^{**}}$	1.455 ± 0.133^{b}	0.000
HDL/LDL		2.338±0.192ª	$2.320{\pm}0.128^a$	1.887±0.312 ^b	1.430 ± 0.095^{b}	0.004
HDL/Cholesterol		1.361±0.113 ^a	0.969±0.0363 ^{b*}	1.406±0.110 ^a	1.362±0.048 ^a	0.004
Atherogenic factor		0.325±0.106 ^a	0.05±0.067 ^b	0.407±0.112 ^a	0.358±0.047ª	0.011
Atherogenic index of pla	sma	0.033±0.0920 ^b	0.088±0.044 ^b	0.59±0.04 ^a	0.22±0.023 ^b	0.000

The ratio of high-density lipoproteins to low-density lipoproteins in the diabetic ovariectomized group was significantly lower than the surgical control group (P = 0.02). Also, the ratio of these two profiles in the diabetic ovariectomized group was significantly lower than the ovariectomized rats (P = 0.004). The ratio of cholesterol to high-density lipoproteins in ovariectomized rats was lower than in the surgical control group (P = 0.021). Also, this ratio was higher in diabetic rats than ovariectomized rats (P = 0.022). The ratio of cholesterol to high-density lipoproteins was higher in ovariectomized diabetic rats than in ovariectomized rats (P = 0.029) (Table 3). The calculation of the plasma atherogenic index in female diabetic rats was significantly higher than in other groups (P = 0.00). Microscopic studies of the pancreatic tissue of the control group revealed the normal structure of the beta cells of the Langerhans islets. Inducing diabetes with alloxan in rats caused severe damage to beta cells and decreased the number of Langerhans islets and cell death. Pancreatic tissue in the ovariectomy group did not show any pathological changes.

The results of the current study showed that after intraperitoneal injection, alloxan monohydrate with a dose of 150 mg per kilogram of body weight induced diabetes and increased blood sugar in rats under study, which remained until the end of the study period. Also, the blood sugar of ovariectomized diabetic rats was significantly lower than the blood sugar of female diabetic rats. However, the absence of ovaries in female diabetic rats did not increase blood sugar compared to the diabetic group with ovaries. Experimental studies have shown that ovariectomy does not affect the glucose level in streptozotocin-induced diabetic rats, ovariectomy hardly affects glucose metabolism (Herrero et al., 1998), and ovariectomy of diabetic rats increases insulin resistance (Tawfik et al., 2015).

Insulin resistance is a condition in which the cells of the body do not respond properly to insulin, and as a result, the absorption of glucose is disrupted and, secondarily, it causes an increase in blood sugar (Pirola *et al.*, 2004). However, the specific cellular mechanism behind the insulin resistance state and the role of the reduction of female sex hormones on blood glucose are not fully understood (Clegg *et al.*, 2006). The rapid absorption of alloxan by insulin-secreting cells (beta cells of pancreatic islets of Langerhans) and the production of oxygen free radicals cause islet necrosis in the pancreas. Compared to other tissues (such as the liver), beta cells are very sensitive to oxygen free radicals and cause a decrease in plasma insulin concentration. In ovariectomized rats, the ratio of liver weight to body weight was significantly reduced compared to other groups. The reason for this decrease can be irreversible damage such as necrosis and planned

death (Unal et al., 2011). The second reason can be the increase in body weight of ovariectomized rats. Rearchers showed that in ovariectomized rats, food intake, body weight, and intra-abdominal fat accumulation increase, which is reversed by estradiol administration. Also, the lack of estrogen in women after menopause is one of the reasons for increasing visceral fat and insulin resistance and increasing the risk of cardiovascular diseases (Piche et al., 2005).

In the present study, changes in several lipid profiles including triglycerides, high-density lipoproteins, very low-density lipoproteins, the ratio of triglycerides to high-density lipoproteins, the cholesterol ratio to high-density lipoproteins, the atherogenic factor and index Atherogenicity was milder in ovariectomized rats than in diabetic rats. Changes in some lipid profiles including triglycerides of very low-density lipoproteins, the triglycerides ratio to very high-density lipoproteins, and atherogenic index were lower in long-term estrogen-reduced and diabetic rats than in nondepleted rats. Varicotomy was diabetic. The combination of ovariectomy and diabetes induction in rats did not have a significant impact on the weight of rats compared to the surgical control group. The histopathological results revealed that in diabetic rats, the increase in blood sugar is associated with the increase in degeneration and necrosis of liver hepatocytes, which can be due to the decrease in the activity of liver antioxidants. So the activity of serum alanine aminotransferase (ALT), gammaglutamyltransferase (GGT), alkaline phosphatase (ALP), and aspartate aminotransferase (AST) in the non-ovariectomized diabetic group was increased compared to the other groups under study. This finding is consistent with other reports; Chaudhry et al. (2007) reported that alloxan inhibits the antioxidant activity of superoxidase dismutase (SOD) and catalase, and liver dysfunction leads to the leakage of these enzymes from liver cells into the bloodstream.

Diabetic and ovariectomized rats had lower levels of serum albumin than rats of the ovariectomized and surgical control group, which can be justified due to changes in the cytoplasm and nucleus of liver tissue cells and damage to the granular endoplasmic reticulum structures. This is consistent with Nagae *et al.* (1991) studies. In the present study, the serum level of alkaline phosphatase enzyme in diabetic and ovariectomized diabetic rats was significantly higher than that in rats of the surgical and ovariectomized control group. Researchers have shown that increased serum levels of alkaline phosphatase are associated with liver, bone, and kidney disorders (Panteghini & Bais, 2007).

The findings of the current study showed that the induction of diabetes in rats is associated with an increase in the average kidney weight to body weight, an increase in urea, and no change in serum creatinine. The increase in serum creatinine and urea in diabetic rats can be considered due to the increase in protein catabolism and as an indicator of kidney failure (Godsland *et al.*, 1993; D'Eon *et al.*, 2002). In this study, some lipid profiles in the groups under experimental study had significant changes, such that the amount of triglycerides, very low-density lipoproteins, and the triglycerides ratio to high-density lipoproteins in diabetic rats were It was significantly more diabetic than ovariectomized rats, which is a suitable predictive measure for coronary heart disease (Hokanson & Austin, 1996) and one of the main characteristics of insulin resistance and metabolic syndrome (Barzi *et al.*, 2005).

In the present study, atherogenic indices were increased in diabetic rats. Atherogenic indices are predictors of heart diseases and cardiovascular complications that progress with the increase of this index (Frohlich & Dobiásová, 2003).

Conclusion

One of the metabolic diseases that occurs frequently in elderly women is diabetes. In this study, the effects of diabetes on some biochemical parameters and histopathological changes in rats were investigated. The findings showed that the long-term reduction of estrogen in ovariectomized diabetic rats can have beneficial changes in reducing the serum levels of glucose, aspartate aminotransferase, alanine aminotransferase, urea, gammaglutamyl transferase, very low-density lipoprotein, and triglyceride compared to non-ovariectomized diabetic rats.

Acknowledgments: None

Conflict of interest: None

Financial support: None

Ethics statement: None

References

- Alonso-Magdalena, P., Ropero, A. B., Carrera, M. P., Cederroth, C. R., Baquie, M., Gauthier, B. R., Nef, S., Stefani, E., & Nadal, A. (2008). Pancreatic Insulin content regulation by the estrogen receptor ERα. *PloS One*, 3(4), e2069.
- Andersson, B., Mattsson, L. A., Hahn, L., Mårin, P., Lapidus, L., Holm, G., Bengtsson, B. A., & Björntorp, P. (1997). Estrogen replacement therapy decreases hyperandrogenicity and improves Glucose homeostasis and plasma lipids in postmenopausal women with noninsulin-dependent diabetes mellitus. The Journal of Clinical Endocrinology & Metabolism, 82(2), 638-643.
- Bakhshwin, D., Faddladdeen, K. A. J., Ali, S. S., Alsaggaf, S. M., & Ayuob, N. N. (2022). Nanoparticles of Costus speciosus ameliorate Diabetes-induced structural changes in rat prostate through mediating the pro-inflammatory cytokines IL 6, IL1β and TNF-α. Molecules, 27(3), 1027. doi:10.3390/molecules27031027
- Banskota, A. H., Nguyen, N. T., Tezuka, Y., Nobukawa, T., & Kadota, S. (2006). Hypoglycemic effects of the wood of

- Taxus yunnanensis on streptozotocin-induced diabetic rats and its active components. *Phytomedicine*, *13*(1-2), 109-114.
- Barzi, F., Patel, A., Woodward, M., Lawes, C. M., Ohkubo, T., Gu, D., Lam, T. H., Ueshima, H., & Asia Pacific Cohort Studies Collaboration. (2005). A comparison of lipid variables as predictors of cardiovascular disease in the Asia Pacific region. *Annals of Epidemiology*, 15(5), 405-413.
- Behr, G. A., Schnorr, C. E., & Moreira, J. C. F. (2012). Increased blood oxidative stress in experimental menopause rat model: The effects of vitamin a low-dose supplementation upon antioxidant status in bilateral ovariectomized rats. Fundamental & Clinical Pharmacology, 26(2), 235-249.
- Chaudhry, J., Ghosh, N. N., Roy, K., & Chandra, R. (2007).

 Antihyperglycemic effect of a new thiazolidinedione analogue and its role in ameliorating oxidative stress in alloxan-induced Diabetic rats. *Life Sciences*, 80(12), 1135-1142
- Clegg, D. J., Brown, L. M., Woods, S. C., & Benoit, S. C. (2006). Gonadal hormones determine sensitivity to central leptin and insulin. *Diabetes*, 55(4), 978-987.
- D'Eon, T. M., Sharoff, C., Chipkin, S. R., Grow, D., Ruby, B. C., & Braun, B. (2002). Regulation of exercise carbohydrate metabolism by estrogen and progesterone in women. American Journal of Physiology-Endocrinology and Metabolism, 283(5), E1046-E1055.
- Dogru, Y. Z., Nacar, T., Unal, D., Aksak, S., Albayrak, A., Odabasoglu, F., Cetin, N., Karakus, E., Gundogdu, C., Gumustekin, K., et al. (2012). Effects of Diabetes mellitus and postmenopausal period on the lungs of rats. *African Journal of Pharmacy and Pharmacology*, 6(27), 1989-2010.
- Ferrara, A., Karter, A. J., Ackerson, L. M., Liu, J. Y., & Selby, J. V. (2001). Hormone replacement therapy is associated with better glycemic control in women with type 2 diabetes: The Northern California Kaiser Permanente Diabetes registry. *Diabetes Care*, 24(7), 1144-1150.
- Frohlich, J., & Dobiásová, M. (2003). Fractional esterification rate of cholesterol and ratio of triglycerides to HDL-cholesterol are powerful predictors of positive findings on coronary angiography. *Clinical Chemistry*, 49(11), 1873-1880.
- Godsland, I. F., Gangar, K., Walton, C., Cust, M. P., Whitehead, M. I., Wynn, V., & Stevenson, J. C. (1993). Insulin resistance, secretion, and elimination in postmenopausal women receiving oral or transdermal hormone replacement therapy. *Metabolism*, 42(7), 846-853.
- Herrero, S., Calvo, O. M., García-Moreno, C., Martín, E., San Román, J. I., Martín, M., García-Talavera, J. R., Calvo, J. J., & del Pino-Montes, J. (1998). Low bone density with normal bone turnover in ovariectomized and streptozotocininduced diabetic rats. *Calcified Tissue International*, 62, 260-265.
- Hokanson, J. E., & Austin, M. A. (1996). Plasma triglyceride level is a risk factor for cardiovascular disease independent of high-density lipoprotein cholesterol level: A metaanalysis of population-based prospective studies. *Journal of Cardiovascular Risk*, 3(2), 213-219.
- Horton, T. J., Miller, E. K., Glueck, D., & Tench, K. (2002). No

- effect of menstrual cycle phase on glucose kinetics and fuel oxidation during moderate-intensity exercise. *American Journal of Physiology-Endocrinology and Metabolism*, 282(4), E752-E762.
- Ikewuchi, J. C., Ikewuchi, C. C., & Ifeanacho, M. O. (2014).
 Attenuation of salt-loading induced cardiomegaly and dyslipidemia in Wistar rats by aqueous leaf extract of Chromolaena odorata. *Pharmacology & Pharmacy*, 5(02), 160-170.
- Jiménez-Estrada, M., Huerta-Reyes, M., Tavera-Hernández, R., Alvarado-Sansininea, J. J., & Alvarez, A. B. (2021). Contributions from Mexican flora for the treatment of Diabetes mellitus: Molecules of Psacalium decompositum (A. Gray) H. Rob & Brettell. *Molecules*, 26(10), 2892. doi:10.3390/molecules26102892
- Lau, L. H. Y., Nano, J., Cecil, A., Schederecker, F., Rathmann, W., Prehn, C., Zeller, T., Lechner, A., Adamski, J., Peters, A., et al. (2021). Cross-sectional and prospective relationships of endogenous progestogens and estrogens with glucose metabolism in men and women: A KORA F4/FF4 study. BMJ Open Diabetes Research and Care, 9(1), e001951.
- Liebmann, M., Asuaje Pfeifer, M., Grupe, K., & Scherneck, S. (2022). Estradiol (E2) improves glucose-stimulated insulin secretion and stabilizes GDM progression in a prediabetic mouse model. *International Journal of Molecular Sciences*, 23(12), 6693. doi:10.3390/ijms23126693
- Nagae, Y., Miyamoto, M., & Miyamoto, H. (1991). Effect of estrogen replacement on liver function in ovariectomized rats. *The Journal of Toxicological Sciences*, 16(3), 87-100.
- Okada, M., Nomura, S., Ikoma, Y., Yamamoto, E., Ito, T., Mitsui, T., Tamakoshi, K., & Mizutani, S. (2003). Effects of postmenopausal hormone replacement therapy on HbA1c levels. *Diabetes Care*, 26(4), 1088-1092.
- Panteghini, M., & Bais, R. (2007). Enzymes In: Tietz fundamentals of clinical chemistry and Molecular Diagnostics. Burtis CA, Ashwood R, Brun DE, Sawyer BG. 6th ed. Philadelphia: Saunders.
- Piche, M. E., Weisnagel, S. J., Corneau, L., Nadeau, A., Bergeron, J., & Lemieux, S. (2005). Contribution of abdominal visceral obesity and Insulin resistance to the cardiovascular risk profile of postmenopausal women. *Diabetes*, 54(3), 770-777.
- Pirola, L., Johnston, A. M., & Van Obberghen, E. (2004).
 Modulation of insulin action. *Diabetologia*, 47, 170-184.
 Sadeghi, M., Sianati, S., Anaraki, D. K., Ghasemi, M., Paydar, M.

- J., Sharif, B., Mehr, S. E., & Dehpour, A. R. (2009). Study of morphine-induced dependence in gonadectomized male and female mice. *Pharmacology Biochemistry and Behavior*, *91*(4), 604-609.
- Salehi, B., Ata, A., V. Anil Kumar, N., Sharopov, F., Ramírez-Alarcón, K., Ruiz-Ortega, A., Abdulmajid Ayatollahi, S., Valere Tsouh Fokou, P., Kobarfard, F., Amiruddin Zakaria, Z., et al. (2019). Antidiabetic potential of medicinal plants and their active components. *Biomolecules*, 9(10), 551. doi:10.3390/biom9100551
- Sekido, T., Nishio, S. I., Ohkubo, Y., Sekido, K., Kitahara, J., Miyamoto, T., & Komatsu, M. (2019). Repression of Insulin gene transcription by indirect genomic signaling via the estrogen receptor in Pancreatic beta cells. *In Vitro Cellular* & *Developmental Biology-Animal*, 55(4), 226-236. doi:10.1007/s11626-019-00328-5
- Speksnijder, E. M., Ten Noever de Brauw, G. V., Malekzadeh, A., Bisschop, P. H., Stenvers, D. J., & Siegelaar, S. E. (2023). Effect of postmenopausal hormone therapy on Glucose regulation in women with type 1 or type 2 Diabetes: A systematic review and meta-analysis. *Diabetes Care*, 46(10), 1866-1875. doi:10.2337/dc23-0451
- Tanaka, S., Hishiki, M., Ogasawara, J., Sorimachi, E., & Nakayama, M. (2017). The deterioration of the glycemic profile during hormone replacement therapy in a patient with fulminant type 1 Diabetes. *Internal Medicine*, 56(5), 531-534. doi:10.2169/internalmedicine.56.7663
- Tawfik, S. H., Mahmoud, B. F., Saad, M. I., Shehata, M., Kamel, M. A., & Helmy, M. H. (2015). Similar and additive effects of ovariectomy and diabetes on insulin resistance and lipid metabolism. *Biochemistry Research International*, 2015, 1-8.
- Unal, D., Aksak, S., Halici, Z., Sengul, O., Polat, B., Unal, B., & Halici, M. (2011). Effects of diabetes mellitus on the rat liver during the postmenopausal period. *Journal of Molecular Histology*, 42(3), 273-287.
- Warnick, G. R., Knopp, R. H., Fitzpatrick, V., & Branson, L. (1990). Estimating low-density lipoprotein cholesterol by the friedewald equation is adequate for classifying patients on the basis of nationally recommended cutpoints. *Clinical Chemistry*, 36(1), 15-19.
- Zhang, Y., Howard, B. V., Cowan, L. D., Yeh, J., Schaefer, C. F., Wild, R. A., Wang, W., & Lee, E. T. (2002). The effect of estrogen use on levels of glucose and insulin and the risk of type 2 diabetes in American Indian postmenopausal women: The strong heart study. *Diabetes Care*, 25(3), 500-504.