Beneficial Effect of *Proso Millet* in Reducing Oxidative Stress and Osteoporosis in Post-Menopausal Women

Derouiche Samir*, Haddig Nour El-houda, Zerzour Aicha

Received: 14 May 2025 / Received in revised form: 18 August 2025, Accepted: 19 August 2025, Published online: 29 August 2025

Abstract

Finding out how Proso millet aqueous extract affected osteoporosis in postmenopausal women in the Guemar El-Oued area was the aim of this study. Three groups of 32 volunteers' women were chosen; as control and menopausal women have osteoporosis before and after Proso millet aqueous extract treatment during 30 days. Some biochemical and hematological parameters were measured. According to our findings, the osteoporosis patient group had significantly higher levels of WBC, lymphocytes, RBC, HCT, PLT, blood glucose, and serum PAL than the control group, and their serum calcium levels significantly decreased (P<0.01). Additionally, the findings show that, in comparison to the control group, the osteoporosis patients' group had considerably lower levels of GSH, catalase, SOD, and ORAC and significantly greater levels of MDA and vitamin C. Menopausal women who receive treatment with Proso millet aqueous extract have improved biochemical, hematological, and oxidative stress markers as well as a decreased risk of osteoporosis. In conclusion, postmenopausal women with osteoporosis benefit from phytotherapy based on Proso millet in terms of calcium status and oxidative stress.

Keywords: Osteoporosis, Post-menopause, Oxidative stress, *Proso millet*

Introduction

The definitive stop of menstruating and the end of one's ability to reproduce are known as menopause. A consequence of ovarian and hypothalamic-pituitary-ovarian axis dysfunction brought on by aging (Weiss *et al.*, 2004). Additionally, a major global public health problem is osteoporosis, a skeletal disorder marked by

Derouiche Samir*

Department of Cellular and Molecular Biology, Faculty of Natural Sciences and Life, University of El Oued, El-Oued 39000, Algeria.

Laboratory of Biodiversity and Application of Biotechnology in the Agricultural Field, University of El Oued, El-Oued 39000, Algeria.

Haddig Nour El-houda, Zerzour Aicha

Department of Cellular and Molecular Biology, Faculty of Natural Sciences and Life, University of El Oued, El-Oued 39000, Algeria.

*E-mail: dersamebio@gmail.com

weakened bones and a higher chance of injury (Genant et al., 1999). Around 200 million women worldwide suffer from osteoporosis. Even while North America and Europe now have the greatest rates of osteoporosis, as population lifespan increases, emerging nations are predicted to see an increase in this risk (Sözen et al., 2017). One important cause of abnormalities in human metabolism and physiology is oxidative stress, as well as a number of illnesses, according to numerous recent scientific research (Atoussi et al., 2021). The primary cause of a number of diseases (Chetehouna et al., 2020) has been identified as oxidative stress, an aberrant state brought on by an excess of oxidants produced in comparison to antioxidants (Chetehouna et al., 2024). Numerous investigations conducted on animals and in vitro have demonstrated that oxidative stress reduces bone formation by decreasing osteoblast survival and differentiation. However, ROS also stimulate osteoclasts, which improve bone resorption (Domazetovic et al., 2017). Clinical research has also indicated that the pathophysiology of bone loss may include ROS and/or antioxidant systems (Abdollahi, 2005; Chidambaranathan & Culathur, 2022; Patatou et al., 2022; You et al., 2023; Pavlova, 2024). Given these findings, our research aims to investigate how millet extract protects volunteer menopausal women against osteoporosis.

Materials and Methods

Patients and Study Design

This study involved 32 women between the ages of 42 and 55 who were split into two groups: 16 healthy control women with an average age of 49.234±0.34 years, 16 menopausal women with osteoporosis with an average age of 49.972±0.112 years, and the third group, which was determined by supplementing the osteoporosis group with powdered Proso millet grains for four weeks. Women with osteoporosis between the ages of 45 and 55 who voluntarily reside in the Guemar El-Oued region are included in this study, as are about control women who are healthy and free of pathologies. Additionally, all women who use medicines during menopause or who suffer from other acute or chronic diseases are not included in this study (Aloufi *et al.*, 2022; Heimes *et al.*, 2022). The El Oued University Ethics Committee's Department of Cellular and Molecular Biology gave its permission to the research protocol (approval number: 25 EC/DCMB/FNSL/EU2020).

Methods and Laboratory Investigations



Blood is drawn from both groups after a morning fast. Following blood collection, the blood is taken in two different kinds of tubes: One is for assessing oxidative stress and hematological parameters (MDA, GSH, SOD, and CAT), while the other is for anticoagulant (EDTA) tubes. For the purpose of achieving the dosage of the following biochemical parameters: glucose, calcium, iron, PAL, vitamin C, and total antioxidant ORAC, samples are centrifuged in dry tubes for 10 minutes at 3000 rpm. The serum is then recovered.

Analytical Methods

Proso millet grain phytochemical screening uses established procedures to determine the phytochemical substances that are present in the grain (Dhanasekar et al., 2022; Saravanakumar et al., 2022; Ekpo et al., 2023; Eteng et al., 2023). The Slinkard and Singleton method (Boulaares et al., 2024) was used to quantitatively analyze the total phenols in phenolic extracts. The total quantity of flavonoid has been identified using the method outlined by Ahn et al. (2007). The Semi-auto Analyzer Mindray BA-88A was used to analyze serum glucose, calcium, iron, and PAL. Biomaghreb commercial kits were used for the measurements. The hematology auto analyzer (Mindray) was used to do hematological analysis (FNS).

Preparation of Erythrocyte and Leukocyte Samples

After centrifuging the blood EDTA tubes for 10 minutes at 2000 rpm, the plasma is removed. After 30 minutes in the freezer, the EDTA tube cap was dissolved in 50 milliliters of TBS buffer (EDTA 2.92M; tris 1.21M; pH=7). To remove the erythrocyte homogenate, the fluid was centrifuged for 10 minutes at 2500 rpm following incubation. Following erythrocyte separation, the leftover material in the EDTA tube is cleaned and spun for 30 minutes at 2,500 rpm. Until the leukocytes are homogenized and isolated, we repeat the procedure multiple times (Miller *et al.*, 1988).

Oxidative Stress Analyses

MDA was quantified using the TBA reagent in compliance with the protocol described by Sastre et al. (2000). According to Weak and Cory, the quantity of reduced glutathione is determined by calculating the optical density that results from the reduction of dithio-bis-2-nitrobenzoic acid, also referred to as the Ellman reagent, with SH groups found in GSH (Weak & Cory, 1988). Employing a UV/visible spectrophotometer to measure the absorption of H2O2 at 560 nm, one may calculate the catalasetriggered loss of H₂O₂ in the sample in line with the Aebi technique (Aebi, 1984). The spectrophotometric absorption at 560 nm, which is determined utilizing the NBT by the superoxide anion (O₂•) test method of SOD activity, serves as the basis for identifying the existence of SOD (Beauchamp & Fridovich, 1971). Plasma vitamin C is tested using the Jagota and Dani technique (Jagota & Dani, 1982) employing different ascorbic acids and the Folin reagent. The overall antioxidant capacity of the serum, or its ORAC (Oxygen Radical Absorbance Capacity), is determined using the Oyaizu technique, which measures the red blood cells' ability to resist hemolysis caused by free radicals in vitro with plasma.

Statistical Analysis

The Minitab 17 statistical assessment tool was used to compare the group's research involves employing the Student's t-test; and Microsoft Excel 2007 assisted us in creating the tests and histograms. A difference is considered statistically significant if P is less than 0.05.

Results and Discussion

Description of the Study Population

Age, height, weight, body mass index, and blood type are among the general socioeconomic statistics for the two subject groups. According to **Table 1**, there are no statistically significant differences between these indicators at P > 0.05.

Table 1. Socioeconomic description of control and osteoporosis patients.

neter	Control	Patients	P-value
(ys)	49.234±0.341	49.972±0.112	0.114
ight (kg)	66.596±0.243	70.390±0.500	0.080
(cm)	160.76±0.164	160.38±0.130	0.204
ss index	26.01±0.182	27.39±0.108	0.152
A	43.33	20	0.04
В	10	10	0.123
AB	6.67	13	0.002
О	40	57	0.032
	(ys) ight (kg) (cm) ss index A B AB	(ys) 49.234±0.341 ight (kg) 66.596±0.243 i(cm) 160.76±0.164 ss index 26.01±0.182 A 43.33 B 10 AB 6.67	(ys) 49.234±0.341 49.972±0.112 ight (kg) 66.596±0.243 70.390±0.500 (cm) 160.76±0.164 160.38±0.130 ss index 26.01±0.182 27.39±0.108 A 43.33 20 B 10 10 AB 6.67 13

Qualitative and Quantitative Phytochemical Analysis of Proso Millet

According to the findings of phytochemical investigations, the aqueous extract of proso millet is rich in several significant chemical components, including phenolic compounds and flavonoids in high concentrations, as well as reducing sugars, terpenoids, alkaloids, tannins, flavonoids, and saponins (Table 2).

Table 2. Phytochemical essays, Total phenols and flavonoids concentration in *Proso millet* aqueous extract.

Phytochemical	P. millet
Flavonoids	+
Tannins	+
Alkaloids	+
Terpenoids	+
Saponins	+
Reducing compounds	+
Polyphenols (mg of GAE/g of Powder)	7.61±0.54
Flavonoids (mg QE/g of Powder)	0.70±0.01

Biochemical and Hematological Parameters

Regarding biochemical markers, **Table 3** shows that the osteoporosis patients group had considerably lower serum calcium levels (P<0.01) and significantly greater blood glucose and serum PAL activity (P<0.01) than the control group. However, compared to the osteoporosis patients group, the osteoporosis following plant extract therapy group had substantially lower blood glucose levels and serum PAL activity (P<0.001) and increased serum calcium (P0.001>). But there was no discernible change in the serum iron readings.

In the other hand, WBC (P<0.001), lymphocytes (P<0.05), red blood cells (RBC) (P<0.05), HCT (P<0.01), and PLT (P<0.05) were significantly lower in the osteoporosis+PM group than in the control group (Arios-Caro *et al.*, 2022; Rudayni *et al.*, 2022). Additionally, **Table 3's** results for the hematological parameters indicate that the group of osteoporosis patients had a notable rise (P<0.05) in WBC, lymphocytes, RBC, HCT, and PLT levels, but no discernible change in hemoglobin levels.

Table 3. Hematological levels of control and experimental group.

Parameter	Control (n=16)	Osteoporosis (n=16)	Osteoporosis + PM (n=16)
Blood glucose (g/l)	0.81±0.021	0.918±0.028**	0.855±0.023
Serum Calcium (g/l)	74.31±1.08	65.15±2.04**	75.07±1.95°
Serum Iron (g/l)	14.56±1.54	14.94±1.48	14.25±1.11

Serum PAL (U/l)	128.8±11.0	183±14.0**	107.3±15.9***c
White Blood Cells (×10³/μl)	5.669±0.286	7.169±0.56*	6.033±0.217°
Lymphocytes (×10³/μl)	2.042±0.073	2.200±0.195*	1.944±0.120 ^a
Hemoglobin (g/dl)	12.57±0.25	13.06±0.18	13.60±0.18***b
Red Blood Cells (×10 ⁶ /μl)	4.562±0.055	4.752±0.086*	4.518±0.079 ^a
Hematocrite (%)	39.8±0.719	43.256±0.603***	$41.262{\pm}0.498^{*b}$
Platelets (×10 ³ /μl)	136.06±1.86	255.6±16.4***	237.9±13.0***a

Oxidative Stress Parameters

Lipid Peroxidation and Reduced Glutathione Levels

The findings in **Figure 1** demonstrate that, in comparison to the control group, the osteoporosis patients' group had considerably greater levels of MDA in their leukocytes (P<0.05) and serum (P<0.001) and considerably lower levels of GSH in their erythrocytes (P<0.001), leukocytes (P<0.001), and serum (P<0.001). The aqueous extract of Proso millet therapy resulted in a substantial drop (P<0.001) in MDA levels in leukocytes and serum and a substantial boost (P<0.01) in GSH levels in erythrocytes, leukocytes, and serum in comparison to the osteoporosis group (P<0.05).

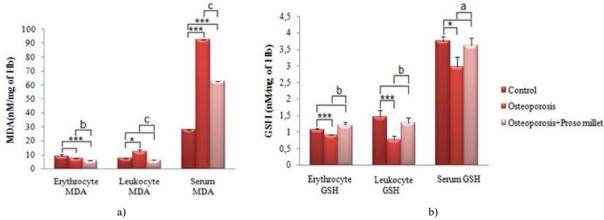


Figure 1. MDA and GSH level of control and experimental group.

Catalase and SOD Activities

The results of one study showed that the osteoporosis patients' group had significantly lower levels of SOD and catalase activities in their serum (P<0.001, P<0.05) and leukocytes (P<0.001) than

the control group (**Figure 2**). On the other hand, the osteoporosis group after plant extract therapy showed noticeably higher levels of catalase and SOD activity in leukocytes (P<0.05, P<0.001) and in serum (P<0.05) than the osteoporosis group.

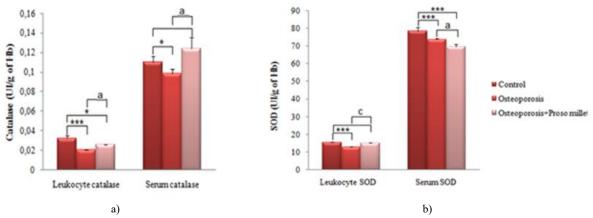


Figure 2. Catalase and SOD activities of control and experimental group.

ORAC and Vitamin C Levels

Following the administration of an aqueous extract of proso millet, the osteoporosis patients' group's serum Oxygen Radical Absorbance Capacity (ORAC) level was considerably greater (P<0.001) than the control group, despite the fact that the ORAC level was considerably lower (P<0.01) than the control group. Additionally, there was not a statistically noteworthy variance in blood vitamin C levels between the osteoporosis group and the control group (P>0.05) as shown in the **Figure 3**.

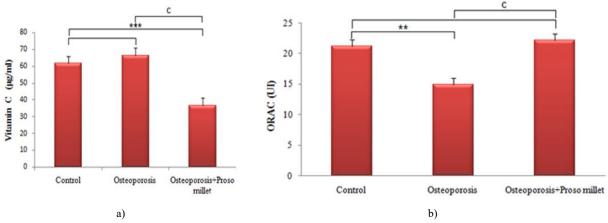


Figure 3. Vitamin C and ORAC levels of control and experimental group.

Phytochemical examination of the aqueous extract of proso millet reveals several significant secondary metabolites, such as reducing sugars, flavonoids, tannins, alkaloids, terpenoids, and saponins. Contains substantial levels of total polyphenols and flavonoids. Among other biological actions, these macromolecules exhibit antioxidant, anti-aging, anti-inflammatory, anti-diabetic, and anticancer properties (Beibalaeva et al., 2022; Osipchuk et al., 2022; Boulaares et al., 2024). Millets' phenols have been discovered to possess a variety of pharmaco-biological properties, such as antimutagenic, anti-oxidant, anti-viral, anti-inflammatory, and aggregation of platelets inhibitory effect (Kumar et al., 2018). Data showed that compared to the control, the patient group's calcium levels significantly decreased and their blood glucose and PAL activity significantly increased, which is consistent with the findings of Tirtha et al. who found that the postmenopausal group had a higher PAL level and a lower serum calcium level (Tirtha et al., 2014). When PAL was raised, all bone mineral density (BMD) readings dramatically dropped. Conversely, bone alkaline phosphatase is a skeletal health indicator that measures bone metabolism (Hailing et al., 2018). The physiology of humans depends on calcium. Is an essential mineral component that contributes to the rigidity of the mature bone's collagen network. One of the main factors contributing to osteoporosis and fracture is inadequate calcium accumulation, which results in a suboptimal bone mass peak and insufficient bone mineralization (DeLucia et al., 2003). Vitamin D may play a role in the establishment of diabetes type II mellitus, according to earlier research. In pancreatic β cells, calcium is transported across the cell membrane and calbindin, a vitamin D-dependent Ca-binding protein, is produced and regulated., are two processes that vitamin D may indirectly influence in relation to insulin release (Liefde et al., 2005). According to the results of our experimental study, patients with osteoporosis who had Proso millet therapy had significantly higher calcium levels than the patient group (Alharbi et al., 2022; Mei & Jiang, 2022; Samaranayake et al., 2024; Menhadji et al., 2024). Although millet, which includes Proso (Panicum miliaceum), gives individuals in many developing countries a lot of calories and protein (Luis et al., 1981), its calcium and phosphorus content is on par with other grains (Burton et al., 1972). According to a study on the mineral content of puffed grains (mg/100 g) (Piłat et al., 2016), Proso millet contains 14.75 mg/100 g of calcium, which could help raise calcium levels. Hematological parameter data indicated that the group of patients with osteoporosis had significantly higher levels of WBC, RBC, HCT, and PLT than the control group. These results provide credence to a potential connection between hematopoiesis and bone metabolism. The development of immature blood cells is known as hematopoiesis. Thus, in the aged, osteoblast activity appears to be directly and intimately linked to bone metabolism and hematopoiesis (Kim et al., 2011; Paspaliaris & Kolios, 2019). Niacin, riboflavin, folic acid, and hydroxycinnamic acid derivatives like p-coumaric and ferulic acid are examples of phenolic compounds that may be responsible for the osteoporosis group's significant decrease in WBC after plant therapy (Derouiche et al., 2022). Si et al. (2014) have shown that niacin lowers vascular inflammatory in both in vitro and in vivo tests by inhibiting the nuclear factor kappa B (NF-κB) signaling pathway. P-coumaric acid and hydroxycinnamic acid have also demonstrated anti-inflammatory qualities (Taofiq et al., 2017). As per the findings of the oxidative stress research, the blood MDA level of the osteoporosis patients was substantially greater than that of the women in the control group (Ilhan et al., 2022; Liu et al., 2022; Mobeen & Dawood, 2022; Ghati et al., 2023). According to a study by Altindag et al., oxidative stress may have contributed to the altered bone metabolism in postmenopausal osteoporosis by increasing the formation of ROS in superoxide forms, as seen by elevated blood MDA levels (Chetehouna et al., 2024). The oxidative stress study's findings demonstrated that the sick group's levels of GSH, catalase, and substantially reduced SOD compared to the control group. A non-enzymatic antioxidant, glutathione (GSH) supports the body's defenses against oxidative stress brought on by free radicals (Chetehouna et al., 2024). Antioxidants play a key role in preventing postmenopausal osteoporosis, according to several studies (Maggio et al., 2003). When osteoporosis patients were compared to the therapy group, the aqueous extract of P. millet's influence on variables related to oxidative stress revealed an extremely substantial rise in leukocyte SOD, serum ORAC, and GSH, as well as an extremely substantial reduction in erythrocyte, leukocyte, and serum MDA levels. Because P. millet extract has anti-inflammatory and antioxidant qualities, it can lessen the effects of oxidation (Habiyaremye et al., 2017). Proso millet contains high concentrations of flavonoids, total polyphenols, hydroxybenzoic acid and its derivatives (vanillic acid, p-hydroxybenzoic acid, and protocatechuic acid), and hydroxycinnamic acid and its derivatives (p-coumaric acid, transferulic acid, cis-ferulic acid, and 5,5'-di ferulic acid) (Chetehouna et al., 2024). During antioxidant processes, proso millet showed a variety of free radical scavenging properties because its phenolic and flavonoid contents predominantly serve as hydrogen donors, reducing free radical, singlet-oxygen quencher's agents, and metal chelating (Kim et al., 2010). The significant decrease in blood vitamin C for osteoporosis + P. millet in comparison to the osteoporosis patient group may be due to insufficient amounts of ascorbat in matured millets (Himanshu et al., 2018). Multiple

epidemiological investigations in postmenopausal women have demonstrated inconsistencies in vitamin C consumption and bone mineral density (Boulaares *et al.*, 2024). Dietary vitamin C and BMD did not independently correlate, according to the Women's Health Initiative Research (Wolf *et al.*, 2005).

Conclusion

This study discovered that the menopause stage linked to osteoporosis is marked by oxidative stress and changes in hematological and biochemical markers, which increases the likelihood of disease complications in women. However, using the plant significantly improves patients' health and lessens disease-related discomfort and problems.

Acknowledgments: None

Conflict of interest: None

Financial support: None

Ethics statement: This study was approved by the Faculty of natural and life Sciences, El-Oued University.

References

Abdollahi, M. (2005). Role of oxidative stress in osteoporosis. *Therapy*, 2(5), 787–796.

Aebi, H. (1984). Catalase in vitro. *Methods in Enzymology, 105*(1), 121–126.

Ahn, M., Kumazawa, S., Usui, Y., Nakamura, J., Matsuka, M., Zhu, F., & Nakayama, T. (2007). Antioxidant activity and constituents of propolis collected in various areas of China. *Food Chemistry*, 101(4), 1383–1392.

Alharbi, I. S., Alharbi, A. S., & Ansari, S. H. (2022). Public awareness and perceptions of orthodontic treatment with Invisalign in Qassim, Saudi Arabia. *Turkish Journal of Public Health Dentistry*, 2(1), 13–18. doi:10.51847/DrpPRdrDrf

Aloufi, F. A., Taleb, M. A., Halawani, R. F., Tammar, A., Mahmood, S., & Rahaman, K. R. (2022). Medical emergency preparedness among dental students: a study from King Abdulaziz University. *Annals of Journal of Dental and Medical Assistance*, 2(2), 14–18. doi:10.51847/jSPCJqmQwe

Arios-Caro, L., López-Martínez, V., Alia-Tejacal, I., Guillén-Sánchez, D., Juárez-López, P., & O, N. B. P. L. (2022). Food attractants and trapping methods for monitoring *Drosophila suzukii* and *Zapronius indianus* (Drosophilidae) in fig orchards. *Entomology Letters*, 2(2), 1–9. doi:10.51847/C3WurKnYg5

Atoussi, O., Chetehouna, S., Boulaares, I., Guemari, I. Y., & Derouiche, S. (2021). Analysis of blood pressure, lipid profile and hematological biomarkers in men addicted to tobacco chewing. Research Journal of Pharmacology and Pharmacodynamics, 13(1), 1–5.

Beauchamp, C., & Fridovich, I. (1971). Superoxide dismutase: improved assays and an assay applicable to acrylamide gels. *Analytical Biochemistry*, *44*(1), 276–287.

- Beibalaeva, S. N., Magomedova, A. T., Kuramagomedova, A. G., Gadzhimagomedov, M. M., Belyaeva, V. A., Rahimova, A. N., & Bondarenko, N. G. (2022). Properties of biologically active compounds and medicinal applications of *Ulomoides* dermestoides beetles. International Journal of Veterinary Research and Allied Sciences, 2(1), 9–14. doi:10.51847/ObPD69eRD4
- Boulaares, I., Derouiche, S., & Guemari, I. Y. (2024). Impact of doxorubicin chemotherapy on oxidative stress status in heart and liver: an experimental study on rats. *Pharma Science Analytical Research Journal*, 6(3), 180067.
- Boulaares, I., Derouiche, S., & Guemari, I. Y. (2024). Protective effect of ObE against doxorubicin-induced immunosuppression and cardiotoxicity in rats. *Research Journal of Pharmacy and Technology*, 17(4), 1839–1843.
- Boulaares, I., Derouiche, S., & Niemann, J. (2024). HPLC-Q-TOF-MS analysis of phenolic compounds, in vitro biological activities and in vivo acute toxicity evaluation of *Ocimum basilicum* L. *Fresenius Environmental Bulletin*, 33(2), 73–82.
- Burton, G. W., Wallace, A. T., & Rachie, K. O. (1972). Chemical composition and nutritive value of pearl millet grain. *Crop Science*, 12(2), 187–188.
- Chetehouna, S., Atoussi, O., Boulaares, I., Guemari, I. Y., & Derouiche, S. (2020). The effect of chronic tobacco smoking on atherogenic index and cardiovascular diseases risk in El-Oued (Algeria) men. Asian Journal of Research in Chemistry, 13(6), 1–5.
- Chetehouna, S., Boulaares, I., Atoussi, O., Guemari, I. Y., & Derouiche, S. (2024). Green nanoparticles as a novel application of nanotechnology in medicine: study of zinc, copper and magnesium nanoparticles. *Recent Pharmacy and Biomedical Sciences*, 8(3), 109–120.
- Chetehouna, S., Derouiche, S., & Atoussi, O. (2024). Identification of new alkaloids from Algerian purslane by HPLC-QTOF-MS and beneficial effect of purslane enriched with zinc on experimental Alzheimer disease in rats. *Current Trends in Biotechnology and Pharmacy*, 18(1), 1595–1607. doi:10.5530/ctbp.2024.1.8
- Chetehouna, S., Derouiche, S., & Reggami, Y., & Boulaares, I. (2024). Sonchus maritimus extract-loaded niosomes bioconjugated by linoleic acid in hepatic encephalopathy induced by high-fructose diet in albino Wistar rats. *Archives of Razi Institute*, 79(1), 194–205. doi:10.32592/ARI.2024.79.1.194
- Chetehouna, S., Derouiche, S., Réggami, Y., Boulaares, I., & Frahtia, A. (2024). Gas chromatography analysis, mineral contents and anti-inflammatory activity of *Sonchus maritimus*. *Tropical Journal of Natural Product Research*, 8(4), 6787–6798.
- Chidambaranathan, A. S., & Culathur, T. (2022). Acupuncture for temporomandibular joint muscular disorder: a prospective clinical assessment of its therapeutic effectiveness. *International Journal of Dental Research and Allied Sciences*, 2(2), 10–15. doi:10.51847/7MWBiwx7jQ
- DeLucia, M. C., Mitnick, M. E., & Carpenter, T. O. (2003).

 Nutritional rickets with normal circulating 25-hydroxyvitamin D: a call for reexamining the role of dietary

- calcium intake in North American infants. *Journal of Clinical Endocrinology & Metabolism*, 88(8), 3539–3545.
- Derouiche, S., Chetehouna, S., & Atoussi, W. (2022). The effects of aqueous leaf extract of *Portulaca oleracea* on haematobiochemical and histopathological changes induced by subchronic aluminium toxicity in male Wistar rats. *Pharmacological Research-Modern Chinese Medicine*, 100101.
- Dhanasekar, P., Rajayyan, J. S., Veerabadiran, Y., Kumar, K. S., Kumar, K. S., & Chinnadurai, N. (2022). Evaluation of alum and purification process of water by coagulation method. Bulletin of Pioneer Research in Medical and Clinical Sciences, 1(2), 1–6. doi:10.51847/R8GyfOmMDh
- Domazetovic, V., Marcucci, G., Iantomasi, T., Brandi, M. L., & Vincenzini, M. T. (2017). Oxidative stress in bone remodeling: role of antioxidants. *Clinical Cases in Mineral* and Bone Metabolism, 14(2), 209–216.
- Ekpo, G. I., Victor, S. E., Eteng, O. E., Ebena, R., Ofonime, N., Umoh, E. U., Uduak, O. L., Ufot, S., & Eyong, U. (2023). Synergistic action of hesperidin and quercetin modulate the efficacy of CCl4-induced nephrotoxicity in rat model. Bulletin of Pioneer Research in Medical and Clinical Sciences, 2(1), 49–57. doi:10.51847/EYAT8W7hWt
- Eteng, O. E., Bassey, N., Eteng, E. I., Okwe, E. P., Ekpo, G., Ekam, V., & Ubana, E. (2023). Effect of vanillic acid and morin on bisphenol S and diethyl phthalate inducenephrotoxicity in male rats. *Bulletin of Pioneer Research in Medical and Clinical Sciences*, 2(1), 25–34. doi:10.51847/JipHmYy6fi
- Genant, H. K., Cooper, C., Poor, G., Reid, I., Ehrlich, G., & Kanis, J. (1999). Interim report and recommendations of the World Health Organization Task-Force for Osteoporosis. *Osteoporosis International*, 10(4), 259–264.
- Ghati, N., Bhatnagar, S., Mahendran, M., Thakur, A., Prasad, K., Kumar, D., Dwivedi, T., Mani, K., Tiwari, P., Gupta, R., et al. (2023). Investigating the role of palliative care education in improving the life quality of women with breast cancer. *Journal of Integrative Nursing and Palliative Care*, 4, 69– 74. doi:10.51847/RFsAtZu8Tv
- Habiyaremye, C., Matanguihan, J. B., Guedes, J. D., Ganjyal, G. M., Whiteman, M. R., Kidwell, K. K., & Murphy, K. M. (2017). Proso millet (*Panicum miliaceum* L.) and its potential for cultivation in the Pacific Northwest, U.S. *Frontiers in Plant Science*, 7(1961), 1–17.
- Hailing, C., Jufen, L., & Qian, W. (2018). Associations between bone-alkaline phosphatase and bone mineral density in adults with and without diabetes. *Journal of List Medicine*, 97(17), 1–7.
- Heimes, D., Mark, N. A., Kuchen, R., Pabst, A., Becker, P., Kyyak, S., Thiem, D. G., Schulze, R., & Kämmerer, P. W. (2022). Awareness and preparedness of Saudi dental trainees on medication-related osteonecrosis of the jaw. *Annals of Journal of Dental and Medical Assistance*, 2(2), 1–9. doi:10.51847/SkHmFpUrKD
- Himanshu, K., Chauhan, M., Sonawane, S. K., & Arya, S. S. (2018). Nutritional and nutraceutical properties of millets. *Clinical Journal of Nutrition and Diet, 1*(1), 1–10.
- İlhan, N., Telli, S., Temel, B., & Aştı, T. (2022). Investigating the

- sexual satisfaction mediating role in the relationship between health literacy and self-care of men with diabetes and women's marital satisfaction. *Journal of Integrative Nursing and Palliative Care, 3*, 19–25. doi:10.51847/sFjL3OLpqg
- Jagota, S. K., & Dani, H. M. (1982). New colorimetric technique for the estimation of vitamin C using Folin phenol reagent. *Analytical Biochemistry*, 127(1), 178–182.
- Kim, H. L., Cho, H. Y., Park, I. Y., Choi, J. M., Kim, M., Jang, H. J., & Hwang, S. M. (2011). The positive association between peripheral blood cell counts and bone mineral density in postmenopausal women. *Yonsei Medical Journal*, 52(5), 739–745.
- Kim, J. S., Hyun, T. K., & Kim, M. J. (2010). Anti-oxidative activities of sorghum, foxtail millet and proso millet extracts. *African Journal of Biotechnology*, 9(18), 2683– 2690.
- Kumar, A., Tomer, V., Kaur, A., Kumar, V., & Gupta, K. (2018).
 Millets: a solution to agrarian and nutritional challenges.
 Agriculture & Food Security, 7(3), 1–15.
- Liefde, I., Klift, M., Laet, C. D., Daele, P., Hofman, A., & Pols, H. (2005). Bone mineral density and fracture risk in type-2 diabetes mellitus: the Rotterdam study. *Osteoporosis International*, 16(12), 1713–1720.
- Liu, M., Tang, Q., Wang, Q., Xie, W., Fan, J., Tang, S., Liu, W., Zhou, Y., & Deng, X. (2022). Studying the sleep quality of first pregnant women in the third trimester of pregnancy and some factors related to it. *Journal of Integrative Nursing and Palliative Care*, 3, 1–6. doi:10.51847/K1PUWsJ24H
- Luis, E. S., Sullivan, T. W., & Nelson, L. A. (1981). Nutrient composition and feeding value of proso millets, sorghum grains, and corn in broiler diets. *Poultry Science*, 61(2), 311–320.
- Maggio, D., Barabani, M., Pierandrei, M., Polidori, M., Catani, M., & Mecocci, P. (2003). Marked decrease in plasma antioxidants in aged osteoporotic women: results of a cross-sectional study. *Clinical Endocrinology & Metabolism*, 88(4), 1523–1527.
- Mei, L., & Jiang, L. (2022). Factors influencing post-treatment relapse in diastema closure. Asian Journal of Periodontics and Orthodontics, 2, 51–55. doi:10.51847/5BKHDdH8UU
- Menhadji, P., Patel, R., Asimakopoulou, K., Quinn, B., Khoshkhounejad, G., Pasha, P., Sanchez, R. G., Ide, M., Kalsi, P., Nibali, L. (2024). The influence of teledentistry on patient satisfaction and treatment results in Saudi Arabia during the COVID-19 pandemic. *Turkish Journal of Public Health Dentistry*, 4(1), 36–43. doi:10.51847/elHXU4OUAa
- Miller, S., Dykes, D., & Polesky, H. (1988). A simple salting out procedure for extracting DNA from human nucleated cells. *Nucleic Acids Research*, *16*(3), 1215.
- Mobeen, T., & Dawood, S. (2022). Studying the effect of perceived social support and mental health on marital burnout in infertile women. *Journal of Integrative Nursing and Palliative Care, 3*, 7–12. doi:10.51847/7DkM3Fkiu3
- Osipchuk, G., Povetkin, S., Shpak, T., Verevkina, M., Bondarenko, N., & Kravchenko, N. (2022). Utilization of biologically active compounds from plant materials in specific physiological states of cows. *International Journal*

- of Veterinary Research and Allied Sciences, 2(2), 21–26. doi:10.51847/wNzvM95prz
- Oyaizu, M. (1986). Studies on products of browning reaction: antioxidant activities of products of browning reaction prepared from glucosamine. *Japanese Journal of Nutrition*, 44(6), 307–315.
- Paspaliaris, V., & Kolios, G. (2019). Stem cells in osteoporosis: from biology to new therapeutic approaches. *Stem Cells International*. doi:10.1155/2019/1730978
- Patatou, A., Iacovou, N., Zaxaria, P., Vasoglou, M., & Vasoglou, G. (2022). Corticotomy-assisted orthodontics: biological basis and clinical applications. *Annals of Orthodontics and Periodontics Special*, 2, 8–13. doi:10.51847/0qGERVSoQm
- Pavlova, Z. (2024). Material properties and clinical performance of 3D-printed complete dentures: a systematic review. Annals of Orthodontics and Periodontics Special, 4, 14–25. doi:10.51847/62izsGtXh4
- Piłat, B., Ogrodowska, D., & Zadernowski, R. (2016). Nutrient content of puffed proso millet (*Panicum miliaceum L.*) and amaranth (*Amaranthus cruentus L.*) grains. *Czech Journal* of Food Sciences, 34(1), 362–369. doi:10.17221/405/2015-CJFS
- Rudayni, H. A., Basher, N. S., Al-Keridis, L. A., Ibrahim, N. A., & Abdelmageed, E. (2022). Exploring the effectiveness of Ocimum basilicum extracts in mosquito larvae management. Entomology Letters, 2(1), 12–18. doi:10.51847/upImR4jWMM
- Samaranayake, L., Tuygunov, N., Schwendicke, F., Osathanon, T., Khurshid, Z., Boymuradov, S. A., & Cahyanto, A. (2024). Artificial intelligence in prosthodontics: transforming diagnosis and treatment planning. Asian Journal of Periodontics and Orthodontics, 4, 9–18. doi:10.51847/nNyZ6VD1da
- Saravanakumar, V., Masi, C., Neme, I., Arjun, K., & Dinakarkumar, Y. (2022). Geographical comparison of phytoconstituents in *Euphorbia hirta*: a pilot study in Ethiopia and India. *Bulletin of Pioneer Research in Medical and Clinical Sciences*, 1(2), 34–41. doi:10.51847/ErNYBrhrFF
- Sastre, J., Pallardó, F. V., Corcia de la Asunción, J., & Viña, J. (2000). Mitochondria, oxidative stress and aging. Free Radical Research, 32(3), 189–198.
- Si, Y., Zhang, Y., Zhao, J., Guo, S., Zhai, L., Yao, S., Sang, H., Yang, N., Song, G., Gu, J., et al. (2014). Niacin inhibits vascular inflammation via downregulating nuclear transcription factor-κB signaling pathway. *Mediators of Inflammation*. doi:10.1155/2014/263786
- Sözen, T., Özişik, L., & Başaran, N. Ç. (2017). An overview and management of osteoporosis. European Journal of Rheumatology, 4(1), 46–56.
- Taofiq, O., González-Paramás, A. M., Barreiro, M. F., & Ferreira, I. (2017). Hydroxycinnamic acids and their derivatives: cosmeceutical significance, challenges and future perspectives, a review. *Molecules*, 22(2), 281.
- Tirtha, B., Koushik, B., Prasenjit, C., & Pallav, S. (2014).
 Correlation of common biochemical markers for bone turnover, serum calcium, and alkaline phosphatase in

- postmenopausal women. Malaysian Journal of Medical Sciences, 21(1), 58-61.
- Weak, B. G., & Cory, J. G. (1988). Ribonucleotide reductase activity and growth of glutathione-depleted leukemia L1210 cells in vitro. *Cancer Letters*, 40(3), 257–264.
- Weiss, G., Skurnick, J. H., Goldsmith, L. T., Santoro, N. F., & Park, S. J. (2004). Menopause and hypothalamic-pituitary sensitivity to estrogen. *JAMA*, 292(24), 2991–2996.
- Wolf, R. L., Cauley, J. A., Pettinger, M., Jackson, R., Lacroix, A., Leboff, M. S., Lewis, C. E., Nevitt, M. C., Simon, J. A.,
- Stone, K. L., et al. (2005). Lack of a relation between vitamin and mineral antioxidants and bone mineral density: results from the Women's Health Initiative. *American Journal of Clinical Nutrition*, 82(3), 581–588.
- You, J. R., Chen, Y. T., Hsieh, C. Y., Chen, S. Y., Lin, T. Y., Shih, J. S., Chen, G. T., Feng, S. W., Peng, T. Y., Wu, C. Y., et al. (2023). Investigating the clinical presentation of oral submucous fibrosis: patterns and progression. *International Journal of Dental Research and Allied Sciences*, 3(2), 9–15. doi:10.51847/SUcIWT7rTw