

Colorectal Cancer Biomarkers Discovery Approach: A Proteomic and Genomic Perspective

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Abstract

Colorectal cancer is a major global health concern, necessitating the identification of biomarkers for early detection, prognosis, and therapeutic targeting. In recent years, proteomic and genomic approaches have revolutionized cancer research, providing valuable insights into the molecular mechanisms underlying CRC. Epidemiological and clinical studies point to a connection between inflammation and development of cancer. Incidences of (CRC) have increased globally during the past decade. In this review, we will discuss the biomarker discovery approach for CRC, which involves a combination of proteomic and genomic perspectives. There are numerous promising biomarkers that must improve life expectancy or quality of life to be evaluated for use in clinical practice. We will cover the important steps such as sample collection, protein extraction, separation, mass spectrometry (MS) analysis, genomic profiling, data analysis, validation, and functional characterization. By utilizing both proteomics and genomics, researchers can identify potential biomarkers for CRC, leading to better diagnostics and personalized treatment options.

Keywords: Colorectal cancer, Biomarkers, Proteomic, Genomic, Gene therapy

Introduction

Cancer is a catastrophic global public health issue, regardless of a country's degree of development (Gari *et al.*, 2021). One million people have diagnosed annually with (CRC), which accounts for 30% of all malignancies.

Incidences of (CRC) have increased globally during the past decade. The third most common adult cancer is CRC, which is also the third leading cause of cancer-related mortality in the United States.

Additionally, the prevalence of CRC is rising in younger people, and by 2030, more people aged 20 to 49 are anticipated to have the disease. CRC prevalence is on the rise in the Middle East, particularly among young people (Coppola *et al.*, 2021). In the

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Arab world, some variations in disease prevalence and epidemiology have also been found. The Arab population has been influenced by Western lifestyles, which have increased the prevalence of CRC and affected younger generations (Guraya, 2018; Makhlof *et al.*, 2021).

Most CRCs are sporadic and are defined by a sequenced carcinogenesis process involving the progressive accumulation of mutations over 10–15 years on average. This long evolution interval permits the successful use of screening, early cancer identification, and treatment of premalignant lesions, decreasing incidence and death (Aghagolzadeh & Radpour, 2016).

Detecting CRC early is the most effective method for reducing cancer mortality. The fecal occult blood test (FOBT), colonoscopy, sigmoidoscopy, and immunological FOBT can detect CRC early enough for effective disease management (Quintero & Salido, 2009; Ettarh, 2012).

Even if there is a potential for an early diagnosis, 20–25% of CRC cases are found at stage IV, when patients have already displayed distant metastases and the 5-year survival probability is less than 10%. For people with early restricted conditions for whom surgical resection is possible, the 5-year survival rate may exceed 90%. Genetic pathways have a major impact on CRC. More than 25% of patients diagnosed with the condition have a family history of it (Corbo *et al.*, 2012; Zygulska & Pierzchalski, 2022).

The large intestine's epithelial cells are the starting point of the prolonged process that leads to CRC development. In reality, these cells lose their normal biological behavior and develop the characteristics of cancer cells as a result of the accumulation of mutations and the subsequent modification in gene function. Depending on the disease's stage, there are three strategies to halt the progression of CRC. The first pertains to cancer or adenoma at an early stage (Gassler *et al.*, 2010).

The last five years have witnessed a significant increase in research aimed at finding biomarkers that can enhance the current diagnostic and prognostic case for CRC screening and therapy (Alkhayat *et al.*, 2021).

Types of Biomarkers



A biomarker is an objectively measurable biological molecule found in bodily fluids or tissues that may be used to identify a pathological state or to indicate whether a biological process is normal or abnormal. Biomarkers can be used to diagnose disease, predict prognosis, and predict pharmacologic responses to therapeutic interventions. Typically, a biomarker must improve life expectancy or quality of life to be evaluated for use in clinical practice (Goossens *et al.*, 2015).

There are three primary categories of biomarkers, depending on the function they serve. Diagnostic biomarkers are likely the most significant biomarkers and are valuable for detecting recurrent disorders. The purpose of prognostic biomarkers is to anticipate the likely course of a disease; they may indicate the aggressiveness and the chance for metastasis. These biomarkers can be used to assess the prognosis of the disease and inform therapy and care decisions. Predictive biomarkers can aid in identifying subpopulations of patients who may benefit from a certain treatment. A predictive biomarker can predict the potential treatment outcomes and can also be employed as a therapy target. It can also signify a "predisposition," or an elevated risk of developing a particular disease. A potential cancer biomarker is any detectable molecular change at the DNA, RNA, protein, or metabolite level in a cancer cell (Atkin, 2003; Diakos *et al.*, 2015).

Yamamoto *et al.* performed liquid chromatography / (MS) on formalin-fixed and paraffin-embedded (FFPE) CRC tissue using a global proteome method, demonstrating greater expression levels of cyclophilin A, annexin A2, and aldolase A in cancer compared to non-cancerous regions (Yamamoto *et al.*, 2016).

Blood-based biomarkers are perhaps the ideal matrix for early diagnosis and surveillance of CRC due to the ease with which non-invasive, low-cost specimens may be collected. Using targeted liquid chromatography-tandem MS (Clarke *et al.*, 2012; Loktionov, 2020).

These results demonstrate the limitations of existing diagnostic screening and the difficulties of generating surrogate markers for early disease identification. Current non-invasive stool screening methods are not sensitive enough to detect precancerous lesions and may miss early-stage CRC. Therefore, a low threshold must be maintained for more intrusive colonoscopies in these patients, and other technologies are necessary to promote early CRC detection.

It is possible to use prognostic biomarkers to predict disease progression, including early recurrence and mortality. KRAS is a member of the RAS proto-oncogene GTPase family, which inhibits cell growth. Mutations in KRAS are associated with a greater likelihood of metastatic CRC recurrence after curative resection, as well as a lower overall survival following hepatic metastasectomy in metastatic CRC (Bonnot & Passot, 2019).

In clinical practice, the primary predictive biomarker is a carcinoembryonic antigen (CEA), a glycoprotein with a high-molecular-weight produced in embryonic tissue and CRCs. This antigen was identified in 1965, but it continues to be the most extensively utilized blood-based biomarker for CRC (Amilca-Seba

et al., 2021; Chen & Ke 2021). Predictive biomarkers are used to personalize therapies based on molecular subtypes. The increasing rise of adjuvant and neoadjuvant therapeutic techniques necessitates the immediate development of predictive biomarkers to guide treatment decisions. An illustration of the significance of predictive biomarkers is the ability of medications to inhibit the epidermal growth factor receptor in patients with KRAS-wild malignancies. The development of this targeting therapy made determining the KRAS status of patients with advanced CRC a prerequisite for determining the efficacy of chemotherapy (Amilca-Seba *et al.*, 2021; Sarkar, 2023).

Genomics and Proteomics

Proteomics encompasses a vast array of techniques used for the large-scale identification, measurement, characterization, and analysis of proteins. The bulk of biomarker discovery research uses quantitative MS-based approaches to identify and validate dysregulated proteins as disease biomarker candidates (Anderson & Anderson, 1998). A genomic biomarker is a detectable DNA or RNA characteristic that serves as an indicator of normal biological activities, pathogenic processes, and/or responsiveness to therapeutic or other interventions (Kim & Hahn, 2007; Bodaghi *et al.*, 2023). A genomic biomarker could, for example, be a measurement of gene expression, function, or regulation (Eltayeb *et al.*, 2022).

Biomarkers Based on Epigenetic Changes for CRC

Epigenetic modifications cause heritable changes in cellular phenotypes and DNA-coded information. These modifications are independent of DNA sequence and susceptible to chromatin-modifying enzymes. Four DNA modifications and sixteen histone modifications have been identified, with cytosine methylations being the most widely described. Complex diseases like cancer, autoimmune disorders, and mental disorders are linked to altered methylation patterns (Schweiger *et al.*, 2013; Zygulska & Pierzchalski, 2022).

In conjunction with posttranscriptional changes of histones, cytosine methylations are arranged in extensive epigenetic silencing areas (LRES). Genes inside these regions are transcriptionally repressed; for instance, a 4-Mb region on chromosome 3p22 containing the MLH1 gene causes MSI-H CRC (Yamashita *et al.*, 2003).

Gene Therapy in CRC

New cancer treatments and tools to analyze genes are leading to a need for dependable biomarkers. Most cancer drugs fail clinical trials, which are expensive and lengthy. To address this, the FDA is prioritizing the use of biomarkers to identify which subtypes of cancer respond best to which treatments. This review covers current trends and challenges in developing effective cancer biomarkers for clinical use (Fearon & Vogelstein, 1990; Jung *et al.*, 2007).

The treatment of CRC is dependent on the TNM staging of cancer, patient health, and curative versus palliative purposes. This includes surgical intervention, chemotherapy, and immunotherapy. The necessity and kind of adjuvant therapy are determined by stage, circumferential resection margin, lymphovascular invasion, perineural invasion, and genotyping (Guetz *et al.*, 2007). 5-FU, commonly used in colon cancer treatment, may harm those with MSI or DPYD. It can improve disease-free survival by 2-4% in stage II CRC, but up to 25% still experience relapse. KRAS wildtype is now being used for better response rates to cetuximab and bevacizumab, while anti-PD-1 drugs treat metastatic CRC. Nivolumab and Ipilimumab have demonstrated efficacy in MSI and mismatch repair defective genotypes, resulting in their approval for patients whose cancer progresses after first-line treatment (Lenz *et al.*, 2022).

Genomics and Biomarker Discovery: Strategies and Their Limitations

Adding anti-EGFR biological medicines to chemotherapy treatment for CRC patients with KRAS mutations can improve survival rates and reduce cancer progression. However, it's unclear if KRAS/BRAF wild-type tumors and KRAS wild-type/BRAF mutant cancers respond differently to anti-EGFR therapy. Studies also conflict with the effectiveness of EGFR-targeted therapies for BRAF-mutant CRC (Garcia-Carbonero *et al.*, 2020).

The P53 gene is crucial for preventing uncontrolled cell growth in cancer. CRC is affected by abnormalities in the TP53 pathway, which can impact treatment effectiveness. More research is needed to determine TP53's potential as a biomarker for CRC (McHugh *et al.*, 2009). MSI status is another indicator of high clinical value. Microsatellites are small DNA sequences that repeat throughout the genome. MSI status is often induced by the inactivation of the four MMR genes (Suzuki *et al.*, 2002). Dysbiosis in the intestinal microbiota may contribute to colorectal carcinogenesis by affecting inflammation, DNA damage, and metabolites involved in tumor progression. This can result from impaired intestinal epithelial barrier function, pro-inflammatory responses, genotoxic biosynthesis, and toxic metabolites produced by pathogens (Tanaka *et al.*, 2010; Goossens *et al.*, 2015).

Genomics Discovery Techniques

Next-generation sequencing (NGS) techniques have changed both genomic and transcriptome analyses. NGS platforms provide deep sequencing, which can detect extremely rare genetic variants, and massively parallel sequencing, which can fast and exhaustively cover the human genome. Different NGS approaches are used for whole-genome sequencing, whole-exome sequencing, targeted sequencing, and RNA-seq. These tools are then used to detect changes in both coding and non-coding genomic regions as well as aberrant dynamics in the transcriptome. High sensitivity and massively parallel sequencing enable quick detection of somatic and germline mutations, propelling NGS to the forefront of cancer biomarker research (McDermott *et al.*, 2013; Marks *et al.*, 2018) MS is the primary enabling tool for proteome discovery. Whatever the case, ionization technique, or performance characteristics, all

mass spectrometers create mass spectra, which plot the mass-to-charge ratio of the observed ions (x-axis) against the measured ion abundance (y-axis) (Fan *et al.*, 2012).

Technology platforms use pattern-based and identity-based techniques to discover proteomic biomarkers. Pattern-based methods generate protein patterns using techniques like SELDI, MALDI, or electrospray. Identity-based methods use LC-MS/MS analysis to identify peptide sequences from differential protein displays like 2D-PAGE. LC-MS/MS-based techniques have proven to be more sensitive, repeatable, and efficient than 2D-PAGE (Mischak *et al.*, 2009). Using methods such as loss of heterozygosity screening and comparative genomic hybridization, the evaluation of the human genome has become extremely efficient (Fanelli *et al.*, 2020). The expression of genes and expressed sequence tags (ESTs) in laboratory and clinical tumor tissues were compared and are now routinely performed using microarray technology (Roos & Byron, 2019).

By amplifying RNA with fluorescent labels and transferring the tagged transcripts on array slides with a large number of oligonucleotides or cDNAs, six thousand genes can be assessed simultaneously. Expression of the fluorescent label indicates the presence and quantity of a certain cDNA transcript in the test population. By combining microarrays with comparative analysis, patterns of gene expression can be detected by logging differences in gene expression (Subramanian *et al.*, 2005).

Proteomics Discovery Techniques

Proteome Marc Wilkins developed the term "proteome" in 1994 by combining the words "protein" and "genome." The proteome encompasses all of the proteins expressed in an organism, tissue, cell, or biological system. Proteomics is the extensive study of all proteins, with an emphasis on their structures and activities (Anderson & Anderson, 1998).

Proteomics is a breakthrough in protein chemistry, focusing on studying the entire proteome as a single analyte for cellular molecular pathways. This approach allows for an accurate representation of the proteome in a given cell state. However, proteome analysis faces challenges such as protein concentrations, detection of post-translational changes (PTMs), and sample complexity (Jungblut *et al.*, 1999). Even though the separation processes vary, the ultimate phase of each strategy is (MS) analysis, which assigns a name to each protein. Several of the most prevalent technologies utilized in CRC research are listed below (Cañas *et al.*, 2006). This area of proteomics employs the methods outlined below and includes (Engwegen *et al.*, 2006):

1. 1-dimensional electrophoresis
2. 2-dimensional electrophoresis
3. In-gel differential electrophoresis
4. Electrophoretic microarrays of proteins
5. Mass Spectrometry

Utilizing Mass Spectrometry

MS has enabled the development of proteomics despite the challenge of detecting low-abundance proteins. Techniques like SILAC, TMT, and iTRAQ have improved sensitivity and allowed for simultaneous analysis and peptide quantification of multiple specimens. TMT LC-MS/MS has high throughput capability and is vital for lab standardization. It has become a prominent technique in identifying cancer biomarkers, resulting in the subclassification of ovarian, breast, and CRCs. CPTAC has successfully used MS to identify cancer-specific proteins and unique protein patterns (Perry *et al.*, 2008).

Antibody-based techniques for targeted proteomics are not the only ones. MS approaches such as Selected Reaction Monitoring (SRM) and Multiple Reaction Monitoring (MRM) are developing as dependable, high-throughput cancer biomarker tests. Specific peptides coming from the protein of interest are identified using a triple quadrupole mass spectrometer and broken down into smaller components, which are then measured to assess protein abundance.

Advances in MS

Through MS-based discovery studies, numerous potential biomarkers for specific diseases have been identified using various technologies. At present, the focus is on creating MS-based MRM scanning techniques to accurately measure the absolute quantity of established proteins in intricate clinical samples. To discover a practical biomarker for therapeutic purposes, customized quantitative proteome profiling methods are necessary, and recent advancements make this increasingly achievable. Nonetheless, the cost of MS instruments and the lack of highly specific antibodies for a significant number of proteins in MS-based biomarker validation methods must be further addressed (Han *et al.*, 2001; Melle *et al.*, 2005; Liu *et al.*, 2006).

Two-Dimensional Gel Electrophoresis

Proteomic biomarker discovery typically uses 2-DE, which separates proteins by charge and size on a gel. However, comparing different gels can be difficult due to slight variations. DIGE minimizes variability, but transferring data between labs is problematic (Ünlü *et al.*, 1997). Secretagoin may indicate abnormal cell differentiation and is being researched in various types of tumors including prostatic adenocarcinoma, pituitary adenomas, and neuroendocrine tumors. It may play a role in the angiogenic activity of human cancer. Further research on living organisms is necessary before it can be used clinically (Alfonso *et al.*, 2005).

In O'Farrell's (1975) original 2DE approach, (O'Farrell, 1975) carrier ampholytes in tube gels are utilized to establish a pH gradient. However, this approach showed limitations in terms of resolution and pH gradient stability. In addition, various sources of variability in 2DE can distort the difference in protein expression, such as (a) analytical variations due to sample treatment, staining procedures, or image acquisition, and (b) biological variations due to the sample's production, processing, and preservation environment. Working with several biological and analytical

replicates helps reduce these differences, but this increases the complexity of the investigation (Friedman *et al.*, 2004).

Matrix-Assisted Laser Desorption/Ionization – Time of Flight

New methods are being used to identify biomarkers, including the Matrix-Assisted Laser Desorption/Ionization – Time of Flight (MALDI-TOF) technology. This involves blending the sample with a matrix molecule that absorbs light at a specific wavelength, then using a laser to transform the sample into gas and eject peptide ions from the surface (de Noo *et al.*, 2006). These ions are separated in a vacuum chamber based on their flight time, and a three-dimensional algorithm is constructed to identify protein clusters. MALDI-TOF has been used to distinguish (CRC) patients from healthy controls, but further validation is needed due to the small sample size and age differences between groups. The technique has also been used to predict metastases in CRC patients, identifying Hsp 27 overexpression as a potential marker for predicting metastatic behavior. These findings are a promising starting point for larger investigations (Liao *et al.*, 2010; Balluff *et al.*, 2011; Kirana *et al.*, 2019).

Surface-Enhanced Laser Desorption Ionization/Time of Flight

SELDI-TOF technology has identified 14 protein peaks that could potentially distinguish RCT responders from nonresponders in rectal cancer patients. These peaks were observed 24-48 hours after the initiation of RCT and remained unchanged at baseline (Seibert *et al.*, 2005; Gemoll *et al.*, 2010).

Sadly, it would be unduly optimistic to believe that targeting these separate proteins will boost patient sensitivity in non-responders, given that a particular tumor's resistance to chemotherapeutic drugs probably involves numerous pathways of resistance. Combination therapy targeting various proteins to sensitize the drug-resistant patient is an aspirational goal for the future of cancer treatment, but the technology is not yet advanced enough to make this a reality (Seibert *et al.*, 2005).

Limitations & Challenges

Gene-expression profiles have limitations as direct biomarkers, as driver genes may not be differentially expressed at the mRNA level, potentially affecting cancer progression. Furthermore, the differentially expressed genes in a signature may not resolve to one or two distinct gene ontological processes, or the pathways to which they map are unclear, limiting their value as mechanistic study guides. In addition, the expression of mRNA is not always proportional to the expression level of the protein, which is the immediate determinant of cellular phenotype. In these instances, gene transcription level may not necessarily have a significant effect on disease. These restrictions should not be interpreted to suggest that genome-wide assessments of protein-coding mRNAs are no longer useful as indicators of dysregulation, which may play a role in illness. Rather, it is important to emphasize that these data are most likely to be useful when combined with all of the essential information we know about the cell.

Conclusion

Gene and protein changes are only part of the complex cellular changes behind colorectal cancer. Proposed gene expression patterns have limited use in predicting the disease. However, systems biology-based methods show promise in identifying markers for various human disorders, despite the challenges of the 'omics revolution. The combination of high-dimensional results from genomes and proteomics, along with legacy data supporting interatomic databases, has the potential to pave the way for more accurate disease classifiers.

Advances in genetics have led to molecular marker assays for colon cancer screening, but current methods fall short of the ideal. FIT and colonoscopy remain the preferred technique. Blood-based screening with the septin9 biomarker has been approved, but its use for precancerous lesions is under review.

Iscoveries in genetics and cancer development have changed how we treat colorectal cancer. Testing for KRAS, BRAF, and MSI status is important for planning therapy. Immunotherapy and liquid biopsies offer new treatment options. However, there are no biomarkers available yet for early diagnosis, treatment, prognosis, or monitoring. Challenges include small study sizes and difficulties in data analysis and interpretation, and the need for confirmation in larger populations.

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References

- Aghagolzadeh, P., & Radpour, R. (2016). New trends in molecular and cellular biomarker discovery for colorectal cancer. *World Journal of Gastroenterology*, 22(25), 5678-5693.
- Alfonso, P., Núñez, A., Madoz-Gurpide, J., Lombardia, L., Sánchez, L., & Casal, J. I. (2005). Proteomic expression analysis of colorectal cancer by two-dimensional differential gel electrophoresis. *Proteomics*, 5(10), 2602-2611. doi:10.1002/pmic.200401196
- Alkhayyat, S., Khojah, M., AlJehan, M., Allali, D., Tayeb, A., Albukhari, S., Qusty, N., Al-Wassia, R., & Baljoon, R. (2021). Awareness of Colorectal Cancer in Saudi Arabia: Cross-Sectional Study. *Pharmacophore*, 12(1), 38-43.
- Amilca-Seba, K., Sabbah, M., Larsen, K., & Denis, A. (2021). Osteopontin as a Regulator of Colorectal Cancer Progression and Its Clinical Applications. *Cancers*, 13(15), 3793. doi:10.3390/cancers13153793
- Anderson, N. L., & Anderson, N. G. (1998). Proteome and proteomics: new technologies, new concepts, and new words. *Electrophoresis*, 19(11), 1853-1861. doi:10.1002/elps.1150191103
- Atkin W. (2003). Options for screening for colorectal cancer. *Scandinavian Journal of Gastroenterology, Supplement*, 38(237), 13-16. doi:10.1080/00855910310001421
- Balluff, B., Schöne, C., Höfler, H., & Walch, A. (2011). MALDI imaging mass spectrometry for direct tissue analysis: technological advancements and recent applications. *Histochemistry and Cell Biology*, 136(3), 227-244. doi:10.1007/s00418-011-0843-x
- Bodaghi, A., Fattahi, N., & Ramazani, A. (2023). Biomarkers: Promising and valuable tools towards diagnosis, prognosis and treatment of Covid-19 and other diseases. *Heliyon*, 9(2), e13323. doi:10.1016/j.heliyon.2023.e13323
- Bonnot, P. E., & Passot, G. (2019). RAS mutation: site of disease and recurrence pattern in colorectal cancer. *Chinese Clinical Oncology*, 8(5), 55. doi:10.21037/cco.2019.08.11
- Cañas, B., López-Ferrer, D., Ramos-Fernández, A., Camafeita, E., & Calvo, E. (2006). Mass spectrometry technologies for proteomics. *Briefings in Functional Genomics & Proteomics*, 4(4), 295-320. doi:10.1093/bfpg/eli002
- Chen, L., & Ke, X. (2021). MMP7 as a potential biomarker of colon cancer and its prognostic value by bioinformatics analysis. *Medicine*, 100(9), e24953. doi:10.1097/MD.00000000000024953
- Clarke, S. J., Karapetis, C. S., Gibbs, P., Pavlakis, N., Desai, J., Michael, M., Tebbutt, N. C., Price, T. J., & Tabernero, J. (2013). Overview of biomarkers in metastatic colorectal cancer: tumor, blood, and patient-related factors. *Critical Reviews in Oncology/hematology*, 85(2), 121-135. doi:10.1016/j.critrevonc.2012.06.001
- Coppola, R., Santo, B., Silipigni, S., & Panasiti, V. (2021). Symmetrical drug-related intertriginous and flexural exanthema and acneiform eruption in a patient with metastatic colorectal cancer treated with cetuximab. *Clinical Cancer Investigation Journal*, 10(6), 331-332.
- Corbo, C., Orrù, S., Gemei, M., Noto, R. D., Mirabelli, P., Imperlini, E., Ruoppolo, M., Vecchio, L. D., & Salvatore, F. (2012). Protein cross-talk in CD133+ colon cancer cells indicates activation of the Wnt pathway and upregulation of SRp20 which is potentially involved in tumorigenicity. *Proteomics*, 12(12), 2045-2059. doi:10.1002/pmic.201100370
- de Noo, M. E., Mertens, B. J., Ozalp, A., Bladergroen, M. R., van der Werff, M. P., van de Velde, C. J., Deelder, A. M., & Tollenaar, R. A. (2006). Detection of colorectal cancer using MALDI-TOF serum protein profiling. *European Journal of Cancer*, 42(8), 1068-1076. doi:10.1016/j.ejca.2005.12.023
- Diakos, K. A., Chua, W., Howell Viive, M., & Clarke, S. (2015). Biomarkers in cancer - biomarkers in metastatic colorectal cancer. *Biomarkers in Cancer*. p. 601-629.
- Eltayeb, L. B., Fallatah, D. I., & Mangi, A. A. (2022). Genomic divergence of Hepatitis C virus towards common prescribed interferon regimens on sustained virologic response (SVR). *Journal of Advanced Pharmacy Education and Research*, 12(3), 59-64.
- Engwegen, J. Y., Gast, M. C., Schellens, J. H., & Beijnen, J. H. (2006). Clinical proteomics: searching for better tumor markers with SELDI-TOF mass spectrometry. *Trends in*

- Pharmacological Sciences*, 27(5), 251-259. doi:10.1016/j.tips.2006.03.003
- Ettarh, R. (2012). Colorectal cancer: it starts and it runs. *Colorectal cancer biology—from genes to tumor*, 1-8.
- Fan, N. J., Gao, C. F., Wang, X. L., Zhao, G., Liu, Q. Y., Zhang, Y. Y., & Cheng, B. G. (2012). Serum peptidome patterns of colorectal cancer based on magnetic bead separation and MALDI-TOF mass spectrometry analysis. *Journal of Biomedicine & Biotechnology*, 2012, 985020. doi:10.1155/2012/985020
- Fanelli, C. A., Depetris, I., Schirripa, M., Brignola, S., BIASON, P., Balistreri, M., Dal Santo, L., Lonardi, S., Munari, G., Loupakis, F., et al. (2020). The heterogeneous clinical and pathological landscapes of metastatic Braf-mutated colorectal cancer. *Cancer Cell International*, 20(1), 30.
- Fearon, E. R., & Vogelstein, B. (1990). A genetic model for colorectal Tumorigenesis. *Cell*, 61(5), 759-767. doi:10.1016/0092-8674(90)90186-i
- Friedman, D. B., Hill, S., Keller, J. W., Merchant, N. B., Levy, S. E., Coffey, R. J., & Caprioli, R. M. (2004). Proteome analysis of human colon cancer by two-dimensional difference gel electrophoresis and mass spectrometry. *Proteomics*, 4(3), 793-811. doi:10.1002/pmic.200300635
- Garcia-Carbonero, N., Martinez-Useros, J., Li, W., Orta, A., Perez, N., Carames, C., Hernandez, T., Moren, I., Serrano, G., & Garcia-Foncillas, J. (2020). KRAS and BRAF mutations as prognostic and predictive biomarkers for standard chemotherapy response in metastatic colorectal cancer: a single institutional study. *Cells*, 9(1), 219. doi:10.3390/cells9010219
- Gari, A., Rawas, G., Mufti, A., & Elemam, O. (2021). BRCA Mutations and PARP Inhibitors in Breast and/or Ovarian Cancer Patients. *International Journal of Pharmaceutical Research and Allied Sciences*, 10(3), 33-49.
- Gassler, C., Kaemmerer, E., & Reinartz, N. K. (2010). Modifier-concept of colorectal carcinogenesis: lipidomics as a technical tool in pathway analysis. *World Journal Gastroenterology*, 16(15), 1820-1827.
- Gemoll, T., Roblick, U. J., Auer, G., Jörnvall, H., & Habermann, J. K. (2010). SELDI-TOF serum proteomics and colorectal cancer: a current overview. *Archives of Physiology and Biochemistry*, 116(4-5), 188-196. doi:10.3109/13813455.2010.495130
- Goossens, N., Nakagawa, S., Sun, X., & Hoshida, Y. (2015). Cancer biomarker discovery and validation. *Translational Cancer Research*, 4(3), 256-269.
- Guetz, P., Cucherousset, J., Benamoun, M., Lagorce, C., Sastre, X., Le Toumelin, P., Uzzan, B., Perret G., Morere, J. F., Breau, J., et al. (2007). Microsatellite instability and sensitivity to FOLFOX treatment in metastatic colorectal cancer. *Anticancer Research*, 27(4C), 2715-2719.
- Guraya, S. Y. (2018). The prevalence and evolving risk factors for colorectal cancer in the Arab world. *Biomedical and Pharmacology Journal*, 11(4), 1773-1780.
- Han, D. K., Eng, J., Zhou, H., & Aebersold, R. (2001). Quantitative profiling of differentiation-induced microsomal proteins using isotope-coded affinity tags and mass spectrometry. *Nature Biotechnology*, 19(10), 946-951. doi:10.1038/nbt1001-946
- Jung, J. J., Jeung, H. C., Lee, J. O., Kim, T. S., Chung, H. C., & Rha, S. Y. (2007). Putative chemosensitivity predictive genes in colorectal cancer cell lines for anticancer agents. *Oncology Reports*, 18(3), 593-599.
- Jungblut, U., Zeindl-Eberhart, E., Stulik, J., Koupilova, K., Pleissner, K., Otto, A., Müller, E., Sokolowska-Köhler, W., Grabher, G., & Stöffler, G. (1999). From Genome to Proteome: Advances in the Practice and Application of Proteomics - Proteomics in human disease: Cancer, heart and infectious diseases. *Electrophoresis*, 20(10), 2100-2110.
- Kim, S. Y., & Hahn, W. C. (2007). Cancer genomics: integrating form and function. *Carcinogenesis*, 28(7), 1387-1392. doi:10.1093/carcin/bgm086
- Kirana, C., Peng, L., Miller, R., Keating, J. P., Glenn, C., Shi, H., Jordan, T. W., Maddern, G. J., & Stubbs, R. S. (2019). Combination of laser microdissection, 2D-DIGE, and MALDI-TOF MS to identify protein biomarkers to predict colorectal cancer spread. *Clinical Proteomics*, 16(1), 1-3. doi:10.1186/s12014-019-9223-7
- Lenz, J., Van Cutsem, E., Luisa Limon, M., Wong, K., Hendlisz, A., Aglietta, M., García-Alfonso, P., Neyns, B., Luppi, G., Cardin, B., et al. (2022). First-Line Nivolumab Plus Low-Dose Ipilimumab for Microsatellite Instability-High/Mismatch Repair-Deficient Metastatic Colorectal Cancer: The Phase II CheckMate 142 Study. *Journal of Clinical Oncology*, 40(2), 161-170. doi:10.1200/JCO.21.01015
- Liao, C. C., Mehta, A., Ward, N. J., Marsh, S., Arulampalam, T., & Norton, J. D. (2010). Analysis of postoperative changes in serum protein expression profiles from colorectal cancer patients by MALDI-TOF mass spectrometry: a pilot methodological study. *World Journal of Surgical Oncology*, 8, 33. doi:10.1186/1477-7819-8-33
- Liu, X. P., Shen, J., Li, Z. F., Yan, L., & Gu, J. (2006). A serum proteomic pattern for the detection of colorectal adenocarcinoma using surface-enhanced laser desorption and ionization mass spectrometry. *Cancer Investigation*, 24(8), 747-753. doi:10.1080/07357900601063873
- Loktionov, A. (2020). Biomarkers for detecting colorectal cancer non-invasively: DNA, RNA or proteins?. *World Journal of Gastrointestinal Oncology*, 12(2), 124. doi:10.4251/wjgo.v12.i2.124
- Makhlouf, N. A., Abdel-Gawad, M., Mahros, A. M., Lashen, S. A., Zaghoul, M., & Eliwa, A. (2021). Colorectal cancer in the Arab world: A systematic review. *World Journal of Gastrointestinal Oncology*, 13(11), 1791-1798.
- Marks, K. M., West, N. P., Morris, E., & Quirke, P. (2018). Clinicopathological, genomic and immunological factors in colorectal cancer prognosis. *The British Journal of Surgery*, 105(2), e99-e109. doi:10.1002/bjs.10756
- McDermott, J. E., Wang, J., Mitchell, H., Webb-Robertson, B. J., Hafen, R., Ramey, J., & Rodland, K. D. (2013). Challenges in Biomarker Discovery: Combining Expert Insights with Statistical Analysis of Complex Omics Data. *Expert opinion*

- on *Medical Diagnostics*, 7(1), 37-51. doi:10.1517/17530059.2012.718329
- McHugh, S. M., O'Donnell, J., & Gillen, P. (2009). Genomic and oncoprotein advances in detection and treatment of colorectal cancer. *World Journal of Surgical Oncology*, 7, 36. doi:10.1186/1477-7819-7-36
- Melle, C., Osterloh, D., Ernst, G., Schimmel, B., Bleul, A., & von Eggeling, F. (2005). Identification of proteins from colorectal cancer tissue by two-dimensional gel electrophoresis and SELDI mass spectrometry. *International Journal of Molecular Medicine*, 16(1), 11-17.
- Mischak, H., Coon, J. J., Novak, J., Weissinger, E. M., Schanstra, J. P., & Dominiczak, A. F. (2009). Capillary electrophoresis-mass spectrometry as a powerful tool in biomarker discovery and clinical diagnosis: an update of recent developments. *Mass Spectrometry Reviews*, 28(5), 703-724. doi:10.1002/mas.20205
- O'Farrell P. H. (1975). High-resolution two-dimensional electrophoresis of proteins. *Journal of Biological Chemistry*, 250(10), 4007-4021.
- Perry, R. H., Cooks, R. G., & Noll, R. J. (2008). Orbitrap mass spectrometry: instrumentation, ion motion, and applications. *Mass Spectrometry Reviews*, 27(6), 661-699. doi:10.1002/mas.20186
- Quintero, A. Z., & Salido E. G. (2009). Blood tests for early detection of colorectal cancer. *Current Colorectal Cancer Reports*, 6(1), 30-37.
- Roos, A., & Byron, S. A. (2019). Genomics-Enabled Precision Medicine for Cancer. *Cancer Treatment and Research*, 178, 137-169. doi:10.1007/978-3-030-16391-4_5
- Sarkar, S. (2023). Proteogenomic Approaches to Understand Gene Mutations and Protein Structural Alterations in Colon Cancer. *Physiologia*, 3(1), 11-29.
- Schweiger, M. R., Hussong, M., Röhr, C., & Lehrach, H. (2013). Genomics and epigenomics of colorectal cancer. *Wiley interdisciplinary reviews. Systems Biology and Medicine*, 5(2), 205-219. doi:10.1002/wsbm.1206
- Seibert, V., Ebert, M. P., & Buschmann, T. (2005). Advances in clinical cancer proteomics: SELDI-ToF-mass spectrometry and biomarker discovery. *Briefings in Functional Genomics & Proteomics*, 4(1), 16-26. doi:10.1093/bfgp/4.1.16
- Subramanian, A., Tamayo, P., Mootha, V. K., Mukherjee, S., Ebert, B. L., Gillette, M. A., Paulovich, A., Pomeroy, S. L., Golub, T. R., Lander, E. S., et al. (2005). Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proceedings of the National Academy of Sciences of the United States of America*, 102(43), 15545-15550. doi:10.1073/pnas.0506580102
- Suzuki, E., Chen, W., Anbazhagan, R., van Engeland, M., Weijnenberg, M. P., Herman, J. G., & Baylin S. B. (2002). A genomic screen for genes upregulated by demethylation and histone deacetylase inhibition in human colorectal cancer. *Nature Genetics*, 31(2), 141-149.
- Tanaka, T., Tanaka, M., Tanaka, T., & Ishigamori, R. (2010). Biomarkers for colorectal cancer. *International Journal of Molecular Sciences*, 11(9), 3209-3225. doi:10.3390/ijms11093209
- Unlü, M., Morgan, M. E., & Minden, J. S. (1997). Difference gel electrophoresis: a single gel method for detecting changes in protein extracts. *Electrophoresis*, 18(11), 2071-2077. doi:10.1002/elps.1150181133
- Yamamoto, T., Kudo, M., Peng, W. X., Takata, H., Takakura, H., Teduka, K., Fujii, T., Mitamura, K., Taga, A., Uchida, E., et al. (2016). Identification of aldolase A as a potential diagnostic biomarker for colorectal cancer based on proteomic analysis using formalin-fixed paraffin-embedded tissue. *Tumor biology: the journal of the International Society for Oncodevelopmental Biology and Medicine*, 37(10), 13595-13606. doi:10.1007/s13277-016-5275-8
- Yamashita, K., Dai, T., Dai, Y., Yamamoto, F., & Perucho, M. (2003). Genetics supersedes epigenetics in colon cancer phenotype. *Cancer Cell*, 4(2), 121-131. doi:10.1016/s1535-6108(03)00190-9
- Zygulska, A. L., & Pierzchalski, P. (2022). Novel diagnostic biomarkers in colorectal cancer. *International Journal of Molecular Sciences*, 23(2), 852. doi:10.3390/ijms23020852