

Production of Polyhydroxyalkanoates from Microalgae- A Review

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Abstract

Polyhydroxyalkanoates are synthesized by microorganisms, similar to synthetic polymers, and completely biodegradable. Polyhydroxyalkanoates or PHAs can be processed in a similar way to synthetic polymers and are considered substitutes for petroleum-based plastics. In the culture media, bacteria and large amounts of organic carbon sources are responsible for 50% of the total cost of production of commercial PHAs. It is only possible for the PHA to have a greater commercial application in a reduction in the cost of production. The potential for microalgae to provide PHAs at a lower cost has been suggested by several studies, they require relatively little nutrients as well as deriving energy from light and CO₂, i.e., being photoautotrophs. The purpose of this paper is to examine biopolymer composition and properties, to determine whether petroleum-based plastics can be replaced by PHAs. Microalgal studies that stimulate PHA synthesis are still in the early stages. However, it remains evident that microalgae are capable of producing biopolymers at a low cost and can be used to contribute to the environment.

Keywords: Polyhydroxyalkanoate, Microalgae, Bioplastic, Polyhydroxybutyrate, Biopolymer

Introduction

Over the past seven decades, the production of petroleum-derived plastics has gained considerable attention and is now the most widely used material in industrial and household settings, replacing wood, glass, metals, and building materials (Peng *et al.*, 2020). Lightweight, robust, durable, and resistant to degradation, plastic materials are highly desirable (Saratale *et al.*, 2021). Despite its widespread use and numerous applications, nearly all the plastic in the world is thrown away as garbage or detritus (Emamahadi *et al.*, 2020). There is an annual production of approximately 140 million tons of plastic in the world, requiring about 150 million tons of fossil fuels to be processed and generating large quantities of waste whose decomposition takes thousands of years (Onen Cinar *et al.*, 2020). Since microbial degradation of plastics is extremely difficult, they have proven to be serious environmental problems. As plastics disperse over great distances and end up in sediments, they can remain in the earth's crust for centuries (Abalansa *et al.*, 2020). The

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consumption of plastic is expected to increase until 2030 in the coming decades (Cruz *et al.*, 2022). The methods for disposing of plastic materials are challenging (Anjum *et al.*, 2016). Degradation rates are extremely low in landfills. Toxic by-products are generated by incineration. Although recycling is possible, it requires a lot of time and plastic materials might change their properties (Li & Wilkins, 2020). Furthermore, the use of recycled materials is limited by additives like paints and coatings. Because they are biodegradable and do not use fossil fuels, plastics made from synthetic materials can be replaced by biopolymers. At present, however, biodegradable alternatives are still more expensive than petroleum-based polymers, making it a challenge to develop environmentally friendly alternatives for commercial and retail markets (D'Adamo *et al.*, 2021). In addition to their similar material properties to polypropylene, polyhydroxyalkanoates are also completely biodegradable and can be extruded and injected in the same manner (Choi *et al.*, 2020). Composting waste materials derived from them is easy since they degrade relatively fast by soil organisms (Hatti-Kaul *et al.*, 2020). Fermenters are used to produce PHA. Heterotrophic bacteria, including *Cupriavidus necator* and recombinant *Escherichia coli*, are responsible for this process (Costa *et al.*, 2018; Tan *et al.*, 2021). In cultivation processes involving organic sources, such as glucose and mineral salts, approximately half of the total production cost is accounted for by organic carbon sources (Price *et al.*, 2020). The cost of manufacturing PHA as a polymer is higher than that of some widely used petroleum-based plastics, limiting its industrial applications (Balaji *et al.*, 2013). Another way to produce PHA is by using the cellular biomass of cyanobacteria or prokaryotic microalgae (Roja *et al.*, 2019). Phytoplankton produces biomass only by converting light and atmospheric CO₂ into biomass. Obtaining PHAs at a lower cost can be achieved with a type of microorganism called microalgae. The fact that they are photoautotrophic and have minimal nutrient needs lead to them accumulating PHAs through oxygenated photosynthesis. To increase PHA's market competitiveness and processability, improved polymer characteristics are essential, apart from reducing production costs. Hence, verifying the possible industrial applications of the obtained biopolymers is vital, to learn more about their properties (Costa *et al.*, 2018). There have been several studies that evaluated the properties of biopolymers obtained from microalgae in comparison to synthetic polymers, but very few have addressed molecular mechanisms underlying the process of microalgae synthesizing biopolymers (Dang *et al.*, 2022). Taking a general overview of the production methods of PHAs from microalgae, this review will focus on the properties and compositions of biopolymers, to compare them to petroleum-based plastics.

Microbial Biopolymers

Microbial biopolymers are natural polymers produced and catabolized through numerous organisms which might be non-poisonous to the host and feature a few benefits over petroleum-primarily based plastics (Saratale *et al.*, 2021). Due to their capacity for utility and brief decomposition through microorganisms inclusive of bacteria, biopolymers are revolutionary and exciting. In microbial cells below stress, those biopolymers collect as reserve substances (Onen *et al.*, 2020). Biologically derived PHAs have proven enormous potential in several fields inclusive of pharmaceuticals, agriculture, biofuel, medicine, disposable, and chromatography (Chen & Zhang, 2018). In many microorganisms, polyhydroxyalkanoates (PHA) polyesters are produced and accumulated, commonly once they enter the desk-bound section of improvement (Bhola *et al.*, 2021). PHAs consist of intracellular inclusions which could keep carbon, energy, and account for as much as 80% of a mobileular's weight (Roja *et al.*, 2019). When different important factors inclusive of oxygen, phosphorus, or nitrogen are scarce, they're produced inside the mobileular as insoluble cytoplasmic inclusions inside the presence of excessive carbon. Because those polymeric substances do now no longer considerably alter the mobileular's osmotic state, they may be saved at excessive concentrations in the mobileular (Zhang *et al.*, 2018). These inclusions are spherical, with suggest sizes of 0.2-0.7 μ m, and encased in a 2 nm membrane of round 2% protein and 0.5% lipids. A phospholipid layer, polymerases, depolymerases, and nonspecific cytosolic proteins all play a function inside the granules (Hamad *et al.*, 2018). The lipid monolayer is important to avoid changing the amorphous to the crystalline polyester state, which might be a lot stable but could induce cell rupture (Galiano *et al.*, 2018). In their work on bacterial polyhydroxyalkanoate granules, Grage *et al.* confirmed the presence of a phospholipid layer in PHA isolates (Grage *et al.*, 2009). The authors assume that the granules have a polyester core by a physical phenomenon with integrated or connected proteins appreciate the PHA synthase, phasins, depolymerizing enzymes, and restrictive proteins additionally to the monolayer of phospholipids. Notwithstanding the growing facts, the presence of a phospholipid coat *in vivo* has nevertheless been proven. Various findings have forged doubt on the presence of the supermolecule layer *in vivo*, notably from the microscopy investigations, which recommend that the lipid coat could type as a result of associate in nursing experimental error throughout PHA extraction and processing (Obruca *et al.*, 2020).

Thermoplastic polymers composed of linear hydroxyalkanoic acid are called PHAs. As one monomeric unit's carboxyl group bonds to another monomeric unit's hydroxyl group, an ester bond is formed (Ranganadha & Chandrasekhar, 2021). **Figure 1** shows how they are structured with "n" reaching 35,000 monomers. Monomers of PHA have various R groups from hydrogen atoms to methyl tridecyl (Reddy *et al.*, 2019). The alkyl side chains of phosphoric acid contain usually a saturated alkyl group. However, they can also show a variety of chemical structures that include alkylated, unsaturated, aromatic, branched, epoxidized, and substituted alkyl groups. PHA thermoplastics' side chains can

be chemically modified by cross-linking unsaturated bonding (Reddy *et al.*, 2017).

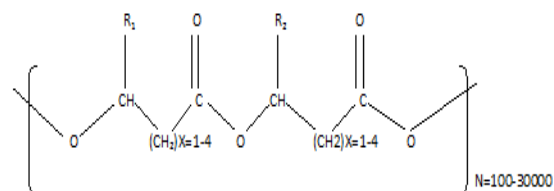


Figure 1. Structure of Polyhydroxyalkanoates with R_1 and R_2 are alkyl groups (C-C)

Essentially, both short-chain and medium-chain hydroxyalkanoic acids have carbon chains of around three to five carbon atoms; medium-chain acids have carbon chains of about six to fourteen, and long-chain acids have carbon chains of more than 14 atoms. PHAs are divided into three groups based on the number of carbon atoms in their carbon chain: short-chain hydroxyalkanoic acids (PHA_{SCL}), medium-chain hydroxyalkanoic acids (PHA_{MCL}), and long-chain hydroxyalkanoic acids (PHA_{LCL}) (Saratale *et al.*, 2021). Among these groups, PHA synthases differ primarily due to their substrate specificity, which allows them to accept three hydroxy acids with a specific carbon length range. Polymers and copolymers can thus be made from different substrates and metabolites depending on the microorganism. PHA_{MCL} is elastomeric and rubber-like, in contrast to PHA_{SCL}, which is comparable to traditional plastics (Ranganadha & Chandrasekhar, 2021). A commonly used commercial form of PHA is PHB (poly-3-hydroxybutyrate). Bacterial bioplastics are generally more expensive than petroleum-based polymers such as polyethylene, polypropylene, and high-density polyethylene which is a limiting factor for their use in the industrial sector. PHA manufacturing costs are often increased due to the high cost of carbon sources (Reddy *et al.*, 2019). More than one hundred microalgae strains are examined for the generation of PHAs. Microalgae use sunshine, carbon dioxide, water, and nutrients in this chemical process cycle to form PHAs collected, purified, and processed in an exceedingly big selection of plastic products. PHAs can be deployed in active microorganism habitats once usage and therefore the microbial degradation of such a compound produce carbonic acid gas and water that microalgae devour (Van-Thuc *et al.*, 2008).

Properties and Composition of PHA's

Thanks to the chemical heterogeneousness of their radicals, PHAs exhibit properties that are just like those of ancient polymers (Ranganadhareddy, 2022). These polymers will vary from onerous and brittle thermoplastics to elastomers, rubbers and adhesives supported the chemical compound makeup (Albuquerque & Malafaia, 2018). They possess a degree of polymerization of up to 30,000 supported by their huge molecular masses. The snap of a polymer rises because the chain length or the amount of comonomers in it grows; therefore, PHAs have varied characteristics looking on their monomeric composition. The molecular mass, thermal characteristics, and crystallinity index are the foremost researched physical parameters of PHAsince they confirm the polymer's processability as shown in

Table 1. A polymer molecular mass and relative molecular mass distribution are crucial qualities for industrial applicability and polymers with molecular lots not up to 4×10^4 Da have poor mechanical properties (Ertan *et al.*, 2021). The molecular mass of those molecules ranges between 2×10^5 to 3×10^5 Da, supported the microorganism species utilized and growth circumstances reminiscent of pH, conditions, culturing modes, and therefore the kind and concentration of the carbon source. The extraction procedure wedged the molecular mass of PHAs as well (Reddy *et al.*, 2019). Laycock *et al.* planned that the molecular mass proportion of biopolymers is connected to their terminal qualities via molecule structural regulations (Laycock *et al.*, 2013). The size of the chemical compound chains determines the PHA characteristics, whereas structural rearrangements are influenced by the degree of polymerization. At room temperature, a polymer's thermal characteristics (T_m – melting temperature, T_g – glass transition temperature) and qualities like crystallinity and crystallization time determine specific mechanical characteristics and are also crucial markers for thermal processing. Depending on the polymer composition, PHAs have a melting temperature ranging from 50 to 180 degrees Celsius and 30 to 70% crystallinity. PHAs possessing crystallinity around 60 and 80 percent, according to Assis *et al.*, are found to be rigid. Accordingly, the polymer lengths of PHAs that are malleable and highly pliable are moderate (30 to 40%) and short (30%). PHA's processing qualities are improved by its reduced degree of crystallinity, enhancing the range of viable industrial uses (Thulasidharan *et al.*, 2021).

Table 1. Physical and thermal properties of different types of PHAs (Costa *et al.*, 2019)

Properties	PHB	PHBV	PP	HDPE
Crystallinity (%)	60-80	30-40	68	60-80
Melting temperature (°C)	37	45	174	125-132
Molecular mass (KDa)	180	164	190	200-600
Breaking strength (%)	4.9	72-87	400	12

The monomeric composition of PHAs is highly reliant on growing conditions and the originating strain, and it is specifically connected to the polymer's characteristics and potential commercial uses (Rojas *et al.*, 2019). Despite the bacteria's ability to acquire more PHAs, the polymers produced are dominated by 4-10 carbon atom monomers (Costa *et al.*, 2018). Large proportions of these monomers give PHAs characteristics that make them unsuitable for industrial use. PHAs have high applicability in various fields such as agriculture, biomedical sciences, and environment, apart from their usual usage in applications such as bag making, fabrics, and so on. With PHA, disposable products, artificial bones, blood vessel substitutes, and heart valve tissue engineering materials can all be made (Thulasidharan *et al.*, 2021).

PHAs Production in Microalgae and Biosynthetic Pathways

According to numerous studies, PHA synthesis by microalgae is known to occur when microbes grow in a nutrient-deficient condition. When microalgae thrive in surroundings deficient in

nitrogen and/or phosphorus, their metabolic pathways are altered to synthesize carbon-rich molecules like PHAs, which operate as a source of energy storage (Mendhulkar & Shetye, 2017). Costa *et al.*, 2018 found that the cyanobacteria *Synechococcus subsalsus* and *Spirulina sp.* LEB-18 are exposed to a nitrogen deficient state; they shift carbon to other metabolic pathways and build biopolymers that can be employed as carbon and energy storage molecules when conditions improve. Various cultivation tactics are used to enhance the biopolymer proportion of PHAs, including the usage of reduced or increased media of specific nutrients, including phosphate, nitrogen, or concurrent deprivation of phosphorus and nitrogen, the inclusion of acetate and propionate, along with higher salinity, restriction of gas exchange, and waste-water usage as sewage, among others. The most frequent metabolic route, which may be seen in a wide spectrum of bacteria that possess an ability to synthesize PHA and fatty acids, is linked to acetyl CoA, a prevalent precursor. The generation of PHAs by microalgae is thought to follow a similar metabolic mechanism. The PHA is derived from acetyl-coenzyme A (acetyl-CoA) by three enzymatic processes as shown in Figure 2 (Li *et al.*, 2021; Ranganadhareddy, 2022).

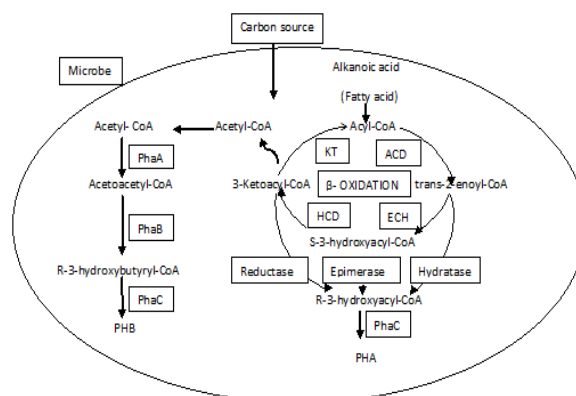


Figure 2. Biosynthetic pathway of Polyhydroxybutyrate in microbes

3- Ketothiolase catalyzes the conversion of two acetyl-CoA molecules as one acetoacetyl-CoA molecule, NADPH-dependent acetoacetyl-CoA reductase catalyzes the reduction of acetoacetyl-CoA to D-3-hydroxybutyryl-CoA and PHA synthase facilitates the association of the D-3-hydroxybutyryl and a PHA molecule through an ester linkage. 3-hydroxy fatty acids are found in β , as well as numerous other PHAs. Between one methyl to thirteen tridecyl carbons make up the pendant group. There are fatty acids having hydroxyl groups at positions 4, 5, and 6 as well as pendant groups carrying substituents or unsaturation, which result in various PHA copolymers and homopolymers (Ranganadha & Chandrasekhar, 2021; Knobloch *et al.*, 2022).

Large scale Production of PHAs from Microalgae

Spirulina sp. was discovered in 1519 in Spain and the study of microalgae and their health benefits increased as a result of the observation of their consumption by Aztecs in the Valley of Mexico, the fact that flamingos survive by eating algae, and flamingos consuming algae as a result of scientific evidence

(Sudhakar *et al.*, 2018). An initial processing plant was established in 1969 to commercialize *Spirulina sp.*, primarily for direct consumption (Cardias *et al.*, 2018). In contrast to the production of microalgae used directly for food, bioplastics produced from microalgae PHAs are not yet commercially available, even though numerous firms are involved in this process, depending on the country, at various stages (Tabet *et al.*, 2018). A typical microalgae provides valuable compositions such as polysaccharides, lipids, proteins, antioxidants, vitamins, pigments, minerals, fatty acids, polyhydroxyalkanoate, micronutrients, etc. A large number of high-value products can be synthesized from the microalgae biomass, including PHAs, which can be used in exceptional commercial applications, microalgae biomass also contains different compounds of high added value, making it possible to commercially acquire highly valuable products (Cardias *et al.*, 2018). According to Singh and Mallick, these demanding situations had to be addressed for making cyanobacterial PHAs a business reality: harvesting of cyanobacterial biomass, drying cyanobacterial biomass, and cyanobacterial mass cultivation. Despite their potential to replace 33% of polymeric materials in the market, PHAs still have a limited market. In upcoming decades, technological improvements in many components in combination with improvements in biological and artificial biology systems will be required to create cyanobacterial species capable of much greater photosynthetic and PHA production (Singh & Mallick, 2017). PHA studies are not only about finding the appropriate biomass and converting it into bioplastic, but also about making it economically sustainable on an industrial scale. The cyanobacteria, in addition to their value as a source of novel and biochemically active herbal products, are additionally being identified as one of the most promising candidates (Kardile & Shirsat, 2020). In the coming years, microalgae production is predicted to keep growing in the global market, resulting in the expansion of the scale of this industry. It is anticipated that biorefineries will convert all biomass compounds into high-value new products identical to those produced by petrochemical refineries.

Conclusion

Sourcing PHAs from microalgae can be viewed as an essential method for lowering polymer acquisition costs and as a result, increasing competitiveness versus synthetic polymers. Microalgae are a viable source for PHAs since these are the only microbes that use photosynthesis to collect PHAs, employing CO₂ and light as their primary sources of energy along with having the low nutritional need for their growth. Microbes consume CO₂ to reduce the warming impact caused by industrial CO₂ emissions. These microbes play an essential role in protecting the environment. In essence, microalgae synthesis of PHAs minimizes the consumption of fossil fuel resources, as well as reduces CO₂ emissions, thereby reducing the environmental impact of the process. Despite, still not knowing how PHAs are produced in microalgae, the growth conditions, species, and extraction methods of these polymers all affect their characteristics, which are crucial for their commercialization. Microalgae-stimulated PHA synthesis is currently in the early

stages of research, and the PHAs produced are not yet commercially available. Regardless, microalgae can create biopolymers at a cheaper cost and play an essential role in the environment.

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References

- Abalansa, S., El Mahrad, B., Vondolia, G. K., Icely, J., & Newton, A. (2020). The marine plastic litter issue: a social-economic analysis. *Sustainability*, *12*(20), 8677. doi:10.3390/su12208677
- Albuquerque, P. B., & Malafaia, C. B. (2018). Perspectives on the production, structural characteristics and potential applications of bioplastics derived from polyhydroxyalkanoates. *International Journal of Biological Macromolecules*, *107*, 615-625. doi:10.1016/j.ijbiomac.2017.09.026
- Anjum, A., Zuber, M., Zia, K. M., Noreen, A., Anjum, M. N., & Tabasum, S. (2016). Microbial production of polyhydroxyalkanoates (PHAs) and its copolymers: a review of recent advancements. *International Journal of Biological Macromolecules*, *89*, 161-174. doi:10.1016/j.ijbiomac.2016.04.069
- Balaji, S., Gopi, K., & Muthuvelan, B. (2013). A review on production of poly β hydroxybutyrate from cyanobacteria for the production of bio plastics. *Algal Research*, *2*(3), 278-285. doi:10.1016/j.algal.2013.03.002
- Bhola, S., Arora, K., Kulshrestha, S., Mehariya, S., Bhatia, R. K., Kaur, P., & Kumar, P. (2021). Established and emerging producers of PHA: Redefining the possibility. *Applied Biochemistry and Biotechnology*, *193*(11), 3812-3854. doi:10.1007/s12010-021-03626-5
- Cardias, B. B., de Moraes, M. G., & Costa, J. A. V. (2018). CO₂ conversion by the integration of biological and chemical methods: *Spirulina sp.* LEB 18 cultivation with diethanolamine and potassium carbonate addition. *Bioresource Technology*, *267*, 77-83. doi:10.1016/j.biortech.2018.07.031
- Chen, G. Q., & Zhang, J. (2018). Microbial polyhydroxyalkanoates as medical implant biomaterials. *Artificial Cells, Nanomedicine, and Biotechnology*, *46*(1), 1-18. doi:10.1080/21691401.2017.1371185
- Choi, S.Y., Rhie, M.N., Kim, H.T., Joo, J.C., Cho, I.J., Son, J., Jo, S.Y., Sohn, Y.J., Baritugo, K. A., Pyo, J., et al. (2020). Metabolic engineering for the synthesis of polyesters: a 100-year journey from polyhydroxyalkanoates to non-natural microbial polyesters. *Metabolic Engineering*, *58*, 47-81. doi:10.1016/j.ymben.2019.05.009

- Costa, S. S., Miranda, A. L., de Moraes, M. G., Costa, J. A. V., & Druzian, J. I. (2019). Microalgae as source of polyhydroxyalkanoates (PHAs)—A review. *International Journal of Biological Macromolecules*, *131*, 536-547. doi:10.1016/j.ijbiomac.2019.03.099.
- Costa, S.S., Miranda, A.L., Andrade, B.B., de Jesus Assis, D., Souza, C.O., de Moraes, M.G., Costa, J.A. V., & Druzian, J.I. (2018). Influence of nitrogen on growth, biomass composition, production, and properties of polyhydroxyalkanoates (PHAs) by microalgae. *International Journal of Biological Macromolecules*, *116*, 552-562. doi:10.1016/j.ijbiomac.2018.05.064
- Cruz, R. A., Oehmen, A., & Reis, M. A. (2022). The impact of biomass withdrawal strategy on the biomass selection and polyhydroxyalkanoates accumulation of mixed microbial cultures. *New Biotechnology*, *66*, 8-15. doi:10.1016/j.nbt.2021.08.004
- D'Adamo, S., Kormelink, R., Martens, D., Barbosa, M. J., & Wijffels, R. H. (2021). Prospects for viruses infecting eukaryotic microalgae in biotechnology. *Biotechnology Advances*, *54*, 107790. doi:10.1016/j.biotechadv.2021.107790
- Dang, B.T., Bui, X.T., Tran, D.P., Ngo, H.H., Nghiem, L.D., Nguyen, P.T., Nguyen, H.H., Lin, C., Lin, K.Y. A., & Varjani, S. (2022). Current application of algae derivatives for bioplastic production: A review. *Bioresource Technology*, *347*, 126698. doi:10.1016/j.biortech.2022.126698
- Emamhadi, M. A., Sarafraz, M., Akbari, M., Fakhri, Y., Linh, N. T. T., & Khaneghah, A. M. (2020). Nanomaterials for food packaging applications: a systematic review. *Food and Chemical Toxicology*, *146*, 111825. doi:10.1016/j.fct.2020.111825.
- Ertan, F., Keskinler, B., & Tanriseven, A. (2021). Exploration of *Cupriavidus necator* ATCC 25207 for the Production of Poly (3-hydroxybutyrate) Using Acid Treated Beet Molasses. *Journal of Polymers and the Environment*, *29*(7), 2111-2125. doi:10.1007/s10924-020-02020-2
- Galiano, F., Briceño, K., Marino, T., Molino, A., Christensen, K. V., & Figoli, A. (2018). Advances in biopolymer-based membrane preparation and applications. *Journal of Membrane Science*, *564*, 562-586. doi:10.1016/j.memsci.2018.07.059
- Grage, K., Jahns, A. C., Parlane, N., Palanisamy, R., Rasiah, I. A., Atwood, J. A., & Rehm, B. H. (2009). Bacterial polyhydroxyalkanoate granules: biogenesis, structure, and potential use as nano-/micro-beads in biotechnological and biomedical applications. *Biomacromolecules*, *10*(4), 660-669. doi:10.1021/bm801394s
- Hamad, K., Kaseem, M., Ayyoob, M., Joo, J., & Deri, F. (2018). Polylactic acid blends: The future of green, light and tough. *Progress in Polymer Science*, *85*, 83-127. doi:10.1016/j.progpolymsci.2018.07.001
- Hatti-Kaul, R., Nilsson, L. J., Zhang, B., Rehnberg, N., & Lundmark, S. (2020). Designing biobased recyclable polymers for plastics. *Trends in Biotechnology*, *38*(1), 50-67. doi:10.1016/j.tibtech.2019.04.011
- Kardile, D., & Shirsat, M. (2020). Synthesis and in vitro evaluation of coupled mercaptobenzimidazole derivatives used as a potent biological agent. *International Journal of Pharmaceutical and Phytopharmacological Research*, *10*(1), 127-133.
- Knobloch, J., Müller, C., & Hildebrandt, J. P. (2022). Expression levels and activities of energy-yielding ATPases in the oligohaline neritid snail *Theodoxus fluviatilis* under changing environmental salinities. *Biology Open*, *11*(2), bio059190. doi:10.1242/bio.059190
- Laycock, B., Halley, P., Pratt, S., Werker, A., & Lant, P. (2013). The chemomechanical properties of microbial polyhydroxyalkanoates. *Progress in Polymer Science*, *38*(3-4), 536-583. doi:10.1016/j.progpolymsci.2012.06.003
- Li, M., & Wilkins, M. R. (2020). Recent advances in polyhydroxyalkanoate production: Feedstocks, strains and process developments. *International Journal of Biological Macromolecules*, *156*, 691-703. doi:10.1016/j.ijbiomac.2020.04.082
- Li, M., Li, K. & Ren, Y. (2021). Nesfatin-1 protects H9c2 cardiomyocytes against cobalt chloride-induced hypoxic injury by modulating the MAPK and Notch1 signaling pathways. *Journal of Biological Research-Thessaloniki*, *28*(21). doi:10.1186/s40709-021-00147-4
- Mendhulkar, V. D., & Shetye, L. A. (2017). Synthesis of biodegradable polymer polyhydroxyalkanoate (PHA) in cyanobacteria *Synechococcus elongatus* under mixotrophic nitrogen-and phosphate-mediated stress conditions. *Industrial Biotechnology*, *13*(2), 85-93. doi:10.1089/ind.2016.0021
- Obruca, S., Sedlacek, P., Slaninova, E., Fritz, I., Daffert, C., Meixner, K., Sedrlova, Z., & Koller, M. (2020). Novel unexpected functions of PHA granules. *Applied Microbiology and Biotechnology*, *104*(11), 4795-4810. doi:10.1007/s00253-020-10568-1
- Onen Cinar, S., Chong, Z. K., Kucuker, M. A., Wiczorek, N., Cengiz, U., & Kuchta, K. (2020). Bioplastic production from microalgae: a review. *International Journal of Environmental Research and Public Health*, *17*(11), 3842. doi:10.3390/ijerph17113842
- Peng, B.Y., Li, Y., Fan, R., Chen, Z., Chen, J., Brandon, A.M., Criddle, C. S., Zhang, Y., & Wu, W.M. (2020). Biodegradation of low-density polyethylene and polystyrene in superworms, larvae of *Zophobas atratus* (Coleoptera: Tenebrionidae): Broad and limited extent depolymerization. *Environmental Pollution*, *266*, 115206. doi:10.1016/j.envpol.2020.115206.
- Price, S., Kuzhiumparambil, U., Pernice, M., & Ralph, P. J. (2020). Cyanobacterial polyhydroxybutyrate for sustainable bioplastic production: critical review and perspectives. *Journal of Environmental Chemical Engineering*, *8*(4), 2213-3437. doi:10.1016/j.jece.2020.104007
- Ranganadhareddy, A. (2022). A Review on Production of Polyhydroxyalkanoates in Microorganisms. *Journal of Biochemical Technology*, *13*(1), 1-6.

- doi:10.51847/Uo3EEbmgID
- Ranganadhareddy, A., & Chandrasekhar, Ch. (2021). Production of polyhydroxybutyrate from marine source- A Review. *Indian Journal of Ecology*, 48(6), 1829-1836.
- Reddy, A. R., Kumar, R. B., & Prabhakar, K. V. (2017). Isolation and Identification of PolyHydroxyButyrate (PHB) producing bacteria from Sewage sample. *Research Journal of Pharmacy and Technology*, 10(4), 1065-1069. doi:10.5958/0974-360X.2017.00193.7
- Reddy, A. R., Peele, K. A., Krupanidhi, S., Prabhakar, K. V., & Venkateswarulu, T. C. (2019). Production of polyhydroxybutyrate from *Acinetobacter nosocomialis* RR20 strain using modified mineral salt medium: a statistical approach. *International Journal of Environment Science and Technology*, 16, 6447-6452. doi:10.1007/s13762-018-2102-3
- Roja, K., Sudhakar, D. R., Anto, S., & Mathimani, T. (2019). Extraction and characterization of polyhydroxyalkanoates from marine green alga and cyanobacteria. *Biocatalysis and Agricultural Biotechnology*, 22, 101358. doi:10.1016/j.bcab.2019.101358
- Saratale, R. G., Cho, S. K., Saratale, G. D., Kadam, A. A., Ghodake, G. S., Kumar, M., Bharagava, R. N., Kumar, G., Kim, D. S., Mulla, S. I., et al. (2021). A comprehensive overview and recent advances on polyhydroxyalkanoates (PHA) production using various organic waste streams. *Bioresource Technology*, 325, 124685. doi:10.1016/j.biortech.2021.124685
- Singh, A. K., & Mallick, N. (2017). Advances in cyanobacterial polyhydroxyalkanoates production. *FEMS Microbiology Letters*, 364(20), 1-13. doi:10.1093/femsle/fnx189
- Sudhakar, K., Mamat, R., Samykano, M., Azmi, W. H., Ishak, W. F. W., & Yusaf, T. (2018). An overview of marine macroalgae as bioresource. *Renewable and Sustainable Energy Reviews*, 91, 165-179. doi:10.1016/j.rser.2018.03.100
- Tabet, A., Boukhari, A., & Nouidjem, Y. (2018). Phenolic content, HPLC analysis and antioxidant activity extract from *Tamarix articulata*. *Journal of Advanced Pharmacy Education & Research*, 8(4), 1-8.
- Tan, D., Wang, Y., Tong, Y., & Chen, G. Q. (2021). Grand challenges for industrializing polyhydroxyalkanoates (PHAs). *Trends in Biotechnology*, 39(9), 953-963. doi:10.1016/j.tibtech.2020.11.010
- Thulasidharan, D., Arumugam, A., & Uppuluri, K. B. (2021). Research and economic perspectives on an integrated biorefinery approach for the simultaneous production of polyhydroxyalkanoates and biohydrogen. *International Journal of Biological Macromolecules*, 193, 1937-1951. doi:10.1016/j.ijbiomac.2021.11.025
- Van-Thuoc, D., Quillaguaman, J., Mamo, G., & Mattiasson, B. (2008). Utilization of agricultural residues for poly (3-hydroxybutyrate) production by *Halomonas boliviensis* LC1. *Journal of Applied Microbiology*, 104(2), 420-428. doi:10.1111/j.1365-2672.2007.03553.x
- Zhang, J., Shishatskaya, E. I., Volova, T. G., da Silva, L. F., & Chen, G. Q. (2018). Polyhydroxyalkanoates (PHA) for therapeutic applications. *Materials Science and Engineering: C*, 86, 144-150. doi:10.1016/j.msec.2017.12.035