Phenolic Compou nds and Antimicrobial Activity of *Ziziphus jujuba* Mill. Fruit from Tlemcen (Algeria)

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Abstract

In this research, the antimicrobial function of the aqueous and organic essences obtained from Ziziphus jujuba pulp fruits was investigated against strains of bacteria and fungi. The RP-HPLC-PDA analysis was observed in the hydro-acetone (EA) and ethyl acetate extracts (Acet1, Acet2) in the presence of gallic acid, quercetin, and rutin. Most extracts present a slight antimicrobial activity with an inhibition zone not exceeding 6.5 mm diameter, except for S. aureus which was the most sensitive strain. The Minimum Inhibitory Concentration (MIC) values ranged between 450 and 110 mg/mL, E. coli and S. aureus are the most sensitive strains for all extracts. No particular antifungal activity of extracts was observed against C. albicans strain. The antimicrobial activity of Z. jujuba pulp fruits has not been reported before, and the present study has revealed that the phenolic compounds identified in jujube fruit extracts cannot provide an important antimicrobial effect to jujube pulp fruits.

Keywords: Ziziphus jujuba Mill., RP-HPLC-PDA analysis, Antimicrobial activity, E. coli

Introduction

Infectious diseases are among the main causes of morbidity and mortality affecting the world, (Ahmad & Beg, 2001; Daneshmand *et al.*, 2013a; Alzaid *et al.*, 2020). Drug-resistant bacteria and fungi

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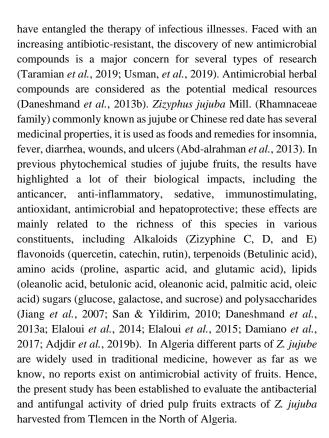
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Materials and Methods

Plant Material

Fruits of *Z. jujuba* were collected in September during the fruit maturation period from Tlemcen city in western Algeria (Latitude: 34° 53′ 24″ Nord, 1° 19′ 12″ Ouest; Altitude: 715m). Botanical identification of the plant was authenticated in the lab of Ecology and Ecosystems Management, Department of Biology, University of Tlemcen, Algeria. In the laboratory, fruits were cleaned and dried at room temperature in the shadow.

Preparation of Plant Extracts

Extracts of Z. jujuba Mill. were obtained by maceration at room temperature: 10 g of dried fruits in 300 ml of solvent: distilled



water for the aqueous extract (Aq); acetone—water and methanolwater mixtures (70/30 v/v) to prepare the extracts EA and EM extracts, respectively. The aqueous (Aq) and acetone-water (EA) extracts underwent fractionation by liquid-liquid extraction using ethyl acetate and n-butanol. The fractions recovered from the acetone-water extract were Acet₂ and n-but₂; those of the aqueous extract were Acet₁ and n-but₁, respectively.

RP-HPLC-PDA Analysis of Phenolic Acids and Flavonoids

Separation and identification of phenolic compounds of the hydroacetone extract EA of *Z. jujube* and the ethyl acetate fractions $Acet_1$ and $Acet_2$ were carried out by RP-HPLC-PDA technique (Adjdir et al., 2019a; El-Haci et al., 2020). Using a binate pump transmission method and an Eclipse ODS Hypersil C18 column (150 mm \times 4.6 μ m), RP-HPLC-PDA investigation of phenolic compositions was carried out on a Perkin Elmar Flexar system. The portable phase included resolvent A- Acetic acid (2%) and B-Acetonitrile. The gradient elution system was: 5 min with 10 % of B; 25 min with 90 % of B and 15 min of linear gradient from 90 % to 100% of B, then, 15 min have included equilibration. The outflow extent was 1 ml/min. The chromatograms were controlled at 280 nm. The compositions' diagnosis and maximum transfers were performed according to their confinement durations as well as correlating with criteria applied.

Assessment of the Antibacterial Function

The Clinical and Laboratory Standards Institute (CLSI) recommended Two approaches that were applied; the disk dispersion and the microdilution approach. Only extracts that have shown important results by the disk-method have been evaluated for their activity by the microdilution method. In this study, five various American Type Culture Collection (ATCC) reference pathogenic bacteria and three fungal strains were utilized. Gramnegative types: *Escherichia coli* ATCC 25933, *Acinetobacter baumanni* ATCC19606, Gram-positive species: *Staphylococcus aureus* ATCC 25923, *Bacillus subtilis* ATCC 6633, *Bacillus cereus* ATCC 11778, and fungal strains *Candida albicans* ATCC 10231, 26790 and IP 444.

The Disk Diffusion Method

Every essence was liquefied in dimethyl sulfoxide (DMSO) in order to obtain 1000 mg/mL ultimate condensation. Gentamicin (Gent $10\mu g/mL$) along with amphotéricin B (AmB $10\mu g/mL$) served as the positive control antibacterial and antifungal. The dimness of the suspensions was regulated at 0.5 McFarland criteria corresponding to 108 CFU/mL. The Petri dishes injected with the suspension regulated at Mueller-Hinton agar by an ear stick. After dehydrating, the antiseptic filter paper disk (6 mm diameter) was subsequently saturated with $10~\mu L$ of every essence. Petri dishes were nurtured at 37 °C, for 18-20 hours. To investigate the sterile or antifungal property, the area of development prevention encompassing the disks was measured.

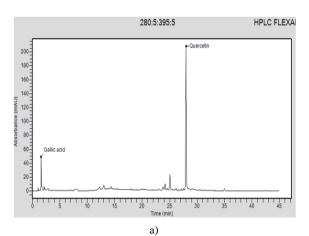
Specification of Minimum Inhibitory Concentration (MIC)

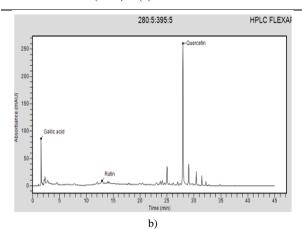
The inoculums got ready and the entire bacterial suspensions were regulated at 0.5 McFarland standard dimness. Sequential subtilizations in Muller Hinton broth were prepared to achieve an ultimate condensation of 5.104CFU /ml by well. In the mentioned experiment, sterile 96 well microplates were utilized. Then, the essences were transmitted to each microplate well, with concentrations ranged from 110 to 450 mg/ml, and the microplates have been nurtured at 35±2 °C, for 16 to 20 hours. The development of the microorganisms in the microdilution wells, as diagnosed by the unaided eye is completely inhibited by the lowest condensation of the essences called the MIC.

Results and Discussion

Recognition of Phenolic Compounds

Chromatographic evaluation by RP-HPLC-PDA indicated in the crude hydro-acetone extract EA and in the ethyl acetate fractions Acet₁ and Acet₂ of Z. jujuba fruits in the presence of gallic acid and quercetin, rutin was only discovered in Acet2 (Figure 1). In a previous study, these extracts revealed a high content in polyphenols, between 50.96 and 94.70 mg gallic acid equivalent per g extract, and in flavonoids from 75.73 to 427.33 mg catechin equivalent per g extract (Adjdir et al., 2019b). In addition, the total phenolic value of fruits methanolic essences from fifteen selected jujube genotypes of Turkey varies between 42 and 40 mg gallic acid equivalent/g DW (Kamiloglu et al., 2009). According to the literature, Gao et al. (2012) have determined a high total phenolic content, in Z. jujube harvested in china, ranged from 275.6 to 541.8 mg of GAE/100 g FW and a dozen phenolic compositions: gallic acid, cinnamic acid, chlorogenic acid, caffeic acid, ferulic acid, ellagic acid, catechin, epicatechin, rutin, and quercetin. Elaloui et al. (2015) have also shown in Z. jujube pulp from Tunisia phenolic compounds especially rutin and chlorogenic acid.





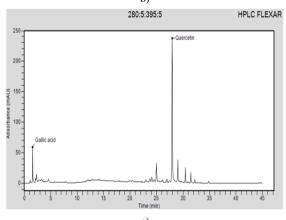


Figure 1. High-performance Liquid Chromatography Profile of Z. Jujube Fruit Extracts
(at 280 nm) a): EA; b): Acet2 and c): Acet1

Antibacterial and Antifungal Activity

The growth inhibition zones of the tested extracts, measured by disk diffusion method, have shown that most extracts present a slight antimicrobial activity with an inhibition zone not exceeding 6.5 mm diameter (Table 1), except for S. aureus which was sensitive to EA (8mm), Acet₂ (7mm), n-but₁ (9mm) and n-but₂ (9mm). The minimum inhibitory concentration results (Table 2) show high MIC values ranged between 450 and 110 mg/mL. E. coli and S. aureus are the most sensitive strains for all extracts. No particular antifungal activity was observed against C. albicans strains (Table 3). On the contrary, several studies have demonstrated the antimicrobial effect of the extracts fruits and other parts from Z. jujube. The ethanolic extract of the fruit is an inhibitor of pediatric infectious strains, S. (MIC=2.26mg/mL), C. albicans (MIC=2.35mg/mL), and A. fumigatus (2.86mg/mL), respectively (Daneshmand et al., 2013b). Snakin-Z, a new cationic peptide (31 amino acids) derived from Z. jujuba fruits has broad-spectrum antimicrobial activity against bacteria, S. aureus (MIC=28.8µg/mL), E. coli (MIC=13.6 µg/mL), and fungi C. albicans (MIC=8.23µg/mL) (Daneshmand et al., 2013a). The hydro-ethanolic extract of seeds inhibits S. aureus (41.25μg/mL; 10mm), E. coli (52.5μg/mL; 17.5mm), and K. pneumonia (42.5µg/mL; 17mm) (Abd-alrahman et al., 2013). Moreover, alphitolic acid a triterpenoid identified from leaves is a significant antibiofilm against S. mutans, a causative agent of human dental caries (Damiano et al., 2017). In the literature, several studies have proved the inhibitory effect of phenolic compounds and flavonoids on microbial strains; it has been reported that Gram-positive bacteria are hypersensitive to herb than Gram-negative due to hydrophobic lipopolysaccharide in the outer membrane that protects various factors (Ferreira et al., 2012; Mezni et al., 2015).

Table 1. Mean Inhibition Zone Diameter (mm) of Ziziphus jujuba Mill. Fruit Extracts on Tested Bacteria by Disk Diffusion Method

		Control (10µg/ disk)						
	Aq	EM	EA	Acet ₁	n-but ₁	Acet ₂	n-but ₂	Gent
Staphylococcus aureus ATCC 25923	$6,5 \pm 0,17$	$6,5 \pm 0,2$	8 ± 0.8	6,5 ±0,6	9 ±1,0	7 ±0,33	9 ±2,5	22
Bacillus cereus ATCC 11778	$6,5\pm0,17$	-	-	-	-	-	6,5 ±0,00	18
Bacillus subtilis ATCC 6633	6,5 ±2,5	-	-	-	-	$6,5 \pm 0,19$	6,5 ±0,6	17
Escherichia coli ATCC 25933	$6,5\pm0,17$	$6,5\pm0,19$	$6,5\pm0,00$	$6,5\pm0,00$	$6,5\pm0,25$	-	$6,5\pm0,17$	18
Acinetobacter baumanii ATCC 19606	$6,5 \pm 0,25$	-	-	-	$6,5 \pm 0,00$	-	-	17

^{(-):} No activity; ND: no determined,

Table 2. The MIC Values (mg/mL) of Z. jujuba Mill. Fruit Extracts against Bacteria

	Extracts							Control
	Aq	EM	EA	Acet ₁	n-but ₁	$Acet_2$	n-but ₂	Gent
Staphylococcus aureus ATCC 25923	450	110	110	230	110	230	110	0,19
Bacillus cereus ATCC 11778	230	-	-	-	-	-	230	0,19
Bacillus subtilis ATCC 6633	230	-	-	-	-	110	230	5,20
Escherichia coli ATCC 25933	450	110	110	110	110	-	110	0,32
Acinetobacter baumanii ATCC 19606	230	-	-	-	110	-	-	0,65

^{(-):} No activity

Control Extracts (1000mg/mL) (10µg/disk) AmB EM EA n-but₁ Acet₂ n-but₂ Αq Acet₁ Candida albicans ATCC10231 32 Candida albicans IP 444 30 Candida albicans ATCC 26790 30

Table 3. Mean Inhibition Zone Diameter (mm) of Z. jujuba Mill. Fruit Extracts on C. albicans Strain by Disk Diffusion Method

(-): No activity

Conclusion

In this study, the antimicrobial activity of *Z. jujuba* fruits pulp from Tlemcen is the first report, these extracts have previously shown *in vitro* an important antioxidant activity related to their higher content on phenolic compounds (Adjdir *et al.*, 2019b). Quercetin, gallic acid, and rutin identified from the pulp fruits have no contribution to the antimicrobial activity of jujube. However, the synergetic effect of these molecules with other phytoconstituents could be primarily responsible for other biological activities of jujube such as the antioxidant effect and the nutritional potential.

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Conflict of Interest: None

Financial Support: None

Ethics Statement: None

References

- Abd-Alrahman, S. H., Salem-Bekhit, M. M., & Elhalwagy, M. E. (2013). Chemical composition and antimicrobial activity of *Ziziphus jujuba* seeds extract. *Journal of Pure and Applied Microbiology*, 7 (Spl. Edn.):379-385.
- Adjdir, S., Benariba, N., El Haci, I. A., Ouffai, K., Bekkara, F. A., & Djaziri, R. (2019a). Antioxidant activity and phenolic compounds identification of Micromeria inodora (Desf.) Benth. from Western Algeria. *Natural Product Research*, 1-
- Adjdir, S., Benariba, N., Laoufi, H., & Djaziri, R. (2019b). Phenolic content and antioxidant activity of *Ziziphus jujuba* Mill. fruit extracts. *Phytothérapie*, 17(2), 74-82.
- Ahmad, I., & Beg, A. Z. (2001). Antimicrobial and phytochemical studies on 45 Indian medicinal plants against multi-drug resistant human pathogens. *Journal of Ethnopharmacology*, 74(2), 113-123.
- Akshita, C., Vijay, B. V., & Praveen, D. (2020). Evaluation of phytochemical screening and antimicrobial efficacy of mesua ferrea and piper cubeba fruit extracts against multidrug-resistant bacteria. *Pharmacophore*, 11(2), 15-20.
- Alzaid, A., Alosaimi, M., Alkahtani, K. F., Alshehri, B. A., Asiri, A. E., Asiri, A. M., Althibait, S. A. S., Aldrees, W. S.,

- Althwaiqub, A. K., Almakhayitah, O. A., et al. (2020). Saudi Parents' Knowledge, Attitudes, and Practices Regarding Antibiotic use for Upper Respiratory Tract Infections in Children. *International Journal of Pharmaceutical Research & Allied Sciences*, 9(1), 115-120.
- Damiano, S., Forino, M., De, A., Vitali, L. A., Lupidi, G., & Taglialatela-Scafati, O. (2017). Antioxidant and antibiofilm activities of secondary metabolites from *Ziziphus jujuba* leaves used for infusion preparation. *Food Chemistry*, 230, 24-29.
- Daneshmand, F., Zare-Zardini, H., & Ebrahimi, L. (2013a). Investigation of the antimicrobial activities of Snakin-Z, a new cationic peptide derived from Zizyphus jujuba fruits. *Natural Product Research*, 27(24), 2292-2296.
- Daneshmand, F., Zare-Zardini, H., Tolueinia, B., Hasani, Z., & Ghanbari, T. (2013b). Crude extract from *Ziziphus jujuba* fruits, a weapon against pediatric infectious disease. *Iranian Journal of Pediatric Hematology and Oncology*, 3(1), 216.
- Elaloui, M., Laamouri, A., Albouchi, A., Cerny, M., Mathieu, C., Vilarem, G., & Hasnaoui, B. (2014). Chemical compositions of the tunisian *Ziziphus jujuba* oil. *Emirates Journal of Food and Agriculture*, 602-608.
- Elaloui, M., Laamouri, A., Fabre, J., Mathieu, C., Vilarem, G., & Hasnaoui, B. (2015). Distribution of free amino acids, polyphenols and sugars of *Ziziphus jujuba* pulps harvested from plants grown in Tunisia. *Natural Product Research*, 29(1), 94-97.
- El-Haci, I. A., Mazari, W., Atik-Bekkara, F., Mouttas-Bendimerad, F., & Hassani, F. (2020). Bioactive compounds from the flower part of Polygonum maritimum L. collected from Algerian coast. *Current Bioactive Compounds*, 16(4), 543-545.
- Ferreira, S., Santos, J., Duarte, A., Duarte, A. P., Queiroz, J. A., & Domingues, F. C. (2012). Screening of antimicrobial activity of *Cistus ladanifer and Arbutus unedo* extracts. *Natural Product Research*, 26(16), 1558-1560.
- Gao, Q. H., Wu, C. S., Yu, J. G., Wang, M., Ma, Y. J., & Li, C. L. (2012). Textural characteristic, antioxidant activity, sugar, organic acid, and phenolic profiles of 10 promising jujube (*Ziziphus jujuba* Mill.) selections. *Journal of Food Science*, 77(11), C1218-C1225.
- Jiang, J. G., Huang, X. J., Chen, J., & Lin, Q. S. (2007). Comparison of the sedative and hypnotic effects of flavonoids, saponins, and polysaccharides extracted from Semen Ziziphus jujube. *Natural Product Research*, 21(4), 310-320.
- Kamiloglu, O., Ercisli, S., Sengül, M., Toplu, C., & Serçe, S. (2009). Total phenolics and antioxidant activity of jujube

- (Zizyphus jujube Mill.) genotypes selected from Turkey. *African Journal of Biotechnology*, 8(2), 303-307.
- Mezni, F., Aouadhi, C., Khouja, M. L., Khaldi, A., & Maaroufi, A.
 (2015). In vitro antimicrobial activity of *Pistacia lentiscus*L. edible oil and phenolic extract. *Natural Product Research*, 29(6), 565-570.
- San, B., & Yildirim, A. N. (2010). Phenolic, alpha-tocopherol, beta-carotene and fatty acid composition of four promising jujube (*Ziziphus jujuba* Miller) selections. *Journal of Food Composition and Analysis*, 23(7), 706-710.
- Taramian, S., Mahfoozi, L., & Soodmand, M. (2019). Antibiotic Resistance Trend of Uropathogens in a Tertiary Teaching Hospital in Rasht, Iran, a Longitudinal Study. *International Journal of Pharmaceutical and Phytopharmacological Research*, 9(1), 98-102.
- Usman, A., Ming, L. C., & Khan, T. M. (2019). Drug-Resistant Sexually Transmitted Infections in Southeast Asia: Regional Challenges to Control. *Archives of Pharmacy Practice*, 10(1), 1-4.